

## Peroxidase Diversity in Hexaploid Wheat *Triticum aestivum* (L.) Thell.

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### Abstract

Seven hexaploid wheat *Triticum aestivum* (L.) Thell. varieties and six  $F_1$ -lines constituted the experimental materials for estimating the peroxidase diversity. The similarity matrix was constructed with Jaccard's coefficient and dendrogram was generated with UPGMA using NTSYSPC version 2.01 software. The differential banding pattern of the  $F_1$ s from their parents may be attributed to reshuffling of genes. Peroxidase polymorphism among wheat varieties represented by their variation in band numbers indicated genotypic variation. Both genetic closeness and distinctness in the molecular level were evident from the results of commonness and distinctness in respect of band numbers and Rm values. Differences in band intensity and band width indicated intervarietal differences in peroxidase activity. The stress tolerant variety C-306 had higher peroxidase activity as compared to other varieties, suggesting that this isozyme synthesis might have taken place as a measure of defensive response to water stress.

**Key words :** Peroxidase, Banding pattern, Diversity, Wheat.

Isoenzymes provide useful evidences in the study of variation between cultivars in terms of intensity of common bands and presence or absences of other bands (1). Genetically the production of isoenzyme of multiple forms or molecular weight is accounted to the allelic variation of the organism. Therefore, isoenzymes of a particular molecular weight can be considered as a direct manifestation of the blue print of the specific gene loci (2). The utility of isoenzymes as genetic marker (3) is generally attributed to their polymorphism, codominance, simple inheritance, simple assay and obliquity in plant tissues or organ (4). Moreover, isoenzymes study may be useful to diversity analysis in plants (5). SDS-PAGE method was found to be useful for studying the peroxidase isoenzymes in wheat (6, 7).

### Methods

Seven hexaploid wheat varieties like K-8027 (V-1), HD-2865 (V-2), DBW-14 (V-3), WUW-234 (V-4), C-306 (V-5), NW-2036 (V-6) and WH-775 (V-7); and six  $F_1$ s like C-2 (V-4  $\times$  V-3), C-3 (V-4 and V-2), C-5 (V-1  $\times$  V-2), C-8 (V-1  $\times$  V-4), C-10 (V-6  $\times$  V-1) and C-12 (V-5  $\times$  V-2) constituted the experimental materials.

To record the electrophoregram of peroxidase the

method followed was that of Kehlar and Allard (8). Based on polyacrylamide gel, bands were scored as present (1) and absent (0) in data sheet to form a [1, 0] matrix. Then data were analyzed and similarity matrix was constructed from binary data with Jaccard's coefficient (9) and dendrogram were generated with unweighted pair group method arithmetic average (UPGMA) algorithm using NTSYSPC-version 2.01 software (10).

### Results and Discussion

The Rm values ranged from 0.16 to 0.64. All bands were found to be polymorphic in nature except band 1 which was present in all varieties and  $F_1$ s but in C-12 it was less intense. Out of seven varieties band three was absent only in V-6 in which band 4 was only present. One more important observation was that the band, which was found in both the parents, was missing in respective  $F_1$ s e.g. band 2, 3 and 5 as in C-8. The differential banding pattern of the  $F_1$ s from their parents is not at all unexpected. This kind of variation might be due to the result of recombination leading to reshuffling of genes.

*Clustering.* All the experimental materials could be grouped into as much as five clusters based on

**Table 1.** Peroxidase banding profile of some wheat varieties along with some selected crosses. Figure in parenthesis indicates total number of band in respective case. +, lightly intense; ++, moderately intense; +++, highly intense band, and -, absence of the band.

Var/F <sub>1</sub> s	Total band	Band 1 (0.16)	Band 2 (0.31)	Band 3 (0.40)	Band 4 (0.46)	Band 5 (0.50)	Band 6 (0.53)	Band 7 (0.64)
V2	4	+++	-	+++	-	-	+++	+++
V3	4	+++	-	+++	-	-	+++	+++
C3 (V4 × V2)	4	+++	-	+++	-	-	+++	+++
V4	4	+++	+	+	-	+++	-	-
C2 (V4 × V3)	4	+++	+++	+++	-	+++	-	-
V1	4	+++	+++	++	-	+++	-	-
C5 (V1 × V2)	4	+++	-	+++	-	+++	-	+++
V7	4	+++	-	+	-	++	-	+
C12 (V5 × V2)	4	+	-	+	-	++	-	+
V5	4	+++	-	++	-	+++	-	++
C10 (V6 × V1)	4	+++	-	-	+	-	-	+
V6	2	+++	-	-	+	-	++	-
C8 (V1 × V4)	3	+++	-	-	+++	-	-	+

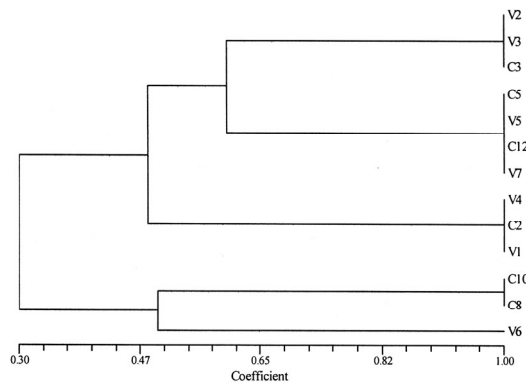
less than 50% Jaccard's similarity coefficient. V-6 was only component of a distinct cluster. C-8 and C-10; V-4, C-2 and V-1; C-5, V-5, C-12 and V-7 and V-2, V-3 and C-3 were the four different distinct clusters (Fig. 1). It was also found that in each cluster, except the cluster having V-6 as sole member; the members were 100% similar to each other. Besides, the similarity coefficient ranged from 0.17 to 0.60 in all other combinations (Table 1). It was cleared that in respect of peroxidase banding profile, the crosses were mostly similar with one of its parent but in some cases showed similarity with other genotypes. The crosses where the effect of both the parents could not be significantly exhibited in respect of peroxidase banding profile as in C-10, C-8 and C-5 (Table 1) might be as a result of recombination leading to the formation of

bands with different molecular weights.

The genotypic variation in respect of band numbers indicated differences among the genotypes. Similar type of peroxidase polymorphism among some rice varieties was observed by Khandelwal et al. (11). This finding also corroborates the previous findings of Roy et al. (12) in grass pea, Philomina and Surendran (5) in neem and Abideen and Vijayakumar (2) in *Acacia* species. Commonness in band numbers and Rm values found in the present experiment indicated their genetic closeness, whereas band number and their relative mobility values when found to be different in two genotypes, indicated their genetic distinctness in the molecular level. Difference in band intensity and band width indicated differences in peroxidase activities. Peroxidase activity is increased in plant tis-

**Table 2.** Jaccard's similarity coefficient for peroxidase banding profile.

Genotypes	V2	V3	C3	V4	C2	V1	C5	V7	C12	V5	C10	V6
V3	1.00											
C3	1.00	1.00										
V4	0.33	0.33	0.33									
C2	0.33	0.33	0.33	1.00								
V1	0.33	0.33	0.33	1.00	1.00							
C5	0.60	0.60	0.60	0.60	0.60	0.60						
V7	0.60	0.60	0.60	0.60	0.60	0.60	1.00					
C12	0.60	0.60	0.60	0.60	0.60	0.60	1.00	1.00				
V5	0.60	0.60	0.60	0.60	0.60	0.60	1.00	1.00	1.00			
C10	0.40	0.40	0.40	0.17	0.17	0.17	0.40	0.40	0.40	0.40		
V6	0.40	0.40	0.40	0.17	0.17	0.17	0.17	0.17	0.17	0.17	0.50	
C8	0.40	0.40	0.40	0.17	0.17	0.17	0.40	0.40	0.40	0.40	1.00	0.50



**Figure 1.** Dendrogram based on peroxidase banding profile.

sues as defensive response to water stress (13). As there is a significant correlation between roor characteristics and water stress condition, the information gathered from peroxidase diversity may help in breeding of wheat varieties with higher peroxidase activity. V-5 (C-306) is a wheat variety generally used as a check for rainfed wheat cultivation (14). As it is a stress tolerant variety, it is expected that peroxidase activity will increase (15). Varietal differences in peroxidase enzyme in rice seedlings grown under stress condition were observed by Pushpam and Rangaswamy (16).

#### References

1. William D. H. M. and A. Mujeeb Kazi. 1992. Isozyme and cytological markers of some *Psathyrostachys juncea* accessions. *Theor. Appl. Genet.* 84 : 534.
2. Abideen Z. Md. and N. K. Vijayakumar. 2002. Isozyme variation in four *Acacia* species. *Indian J. Genet.* 62 : 373—374.
3. Cheniany M., H. Ebrahimzadeh, A. Salimi and V. Niknam. 2007. Isozyme variation in some populations of wild diploid wheats in Iran. *Biochem., Syst. and Ecol.* 35 : 363—371.
4. Simpson M. J. and L. A. Withers. 1986. Characterization of plant genetic resources using isozyme electrophoresis. A guide to the literature, IBPGR, Rome, pp. 258.
5. Philomina D. and C. Surendran. 2003. The application of isozymes to diversity studies in neem (*Azadirachta indica*. A Juss). *Indian J. Genet.* 63 : 93—94.
6. Ju-ZhengChun, Sun- Lan Zhen, Ju- Zc and Sun- LZ. 1997. A comparative study on peroxidase isozyme of K-type and V-type male sterile lines of common wheat and their maintainer lines. *Acta Agriculturae Boreali Sinica* 12 : 7—11.
7. Li- ShuHua Li- SH. 1996. Zymogram analysis of peroxidase isoenzyme of spring wheat varieties in Ningxia. *Ningxia J. Agric. and For. Sci. and Tech.* 2 : 10—13.
8. Kehlar A. L. and R. W. Allard. 1970. Genetics of isozyme variants barley 1 esterase. *Crop Sci.* 10 : 444—448.
9. Jaccard P. 1908. Nouvelles recherches sur la distribution forale. *Bull. Soc. Vaud. Sci. Nat.* 44 : 223—270.
10. Rohlf P. J. 2000. NTSYSpc. Numerical taxonomy and multivariate analysis system, version 2.01. Applied Biostatistics, New York, USA.
11. Khandelwal V., V. Sharma and D. Singh. 2004. Stability for grain yield in sorghum *Sorghum bicolor* (L.) Moench. *Indian J. Genet.* 65 : 53—54.
12. Roy M., N. Mondal and P. K. Das. 2001. Seed protein characterization and isozyme diversity for cultivar identification in grass pea. *Indian J. Genet.* 61 : 246—249.
13. Badiani M., M. D. Biasi, M. Colognola, F. Artemi and M. G. D. Biasi. 1990. Catalase, peroxidase and superoxide dismutase activities in seedling submitted increasing water deficit. *Agrochimica* 34 : 90—102.
14. Chopra R. K. and D. S. Selote. 2007. Acclimation to drought stress generates oxidative stress tolerance in drought-resistant than susceptible wheat cultivar under field conditions. *Environm. and Experim. Bot.* 60 : 276—283.
15. Ashraf M. Y., A. R. Azmi, A. H. Khan and S. A. Ala. 1994. Effect of water stress on total phenols, peroxidase activity and chlorophyll content in wheat (*Triticum aestivum* L.). *Acta Physiol. Planta.* 16 : 185—191.
16. Pushpam R. and S. R. Sree Rangaswamy. 2006. Varietal differences in peroxidase isozyme and protein profiles in rice seedling growing under salinity stress. *Ad. Pl. Sci.* 19 : 313—314.