

Floristic Conservation and Phenological Study on Some Valuable Medicinal Plant Species in Sadhuragiri Hills, Southern Western Ghats of Tamil Nadu, India

Aadhan K., Anand S. P.

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Abstract The document of various periodic behaviors of medicinal plant species or the phenology has great importance because it not only provides knowledge about the plant growth pattern but also provides the inferences on the effect of environment and selective pressure on flowering and fruiting behavior. In this document an attempt has been made to record such data regarding periods of medicinal plants leaf fall, leaf flushing, flowering, fruiting and all. Of the 165 taxa valuable medicinal plants occurring in the southern most hills (Sadhuragiri) of the Southern Western Ghats, Moist deciduous forest (34.5%), Semi evergreen forest (26.06%), Tropical thorn forest (18.18%), Low evergreen forest (16.36%) and Grass land (1.21%) vegetation types. In the study the leaf fall peak period was found in last part of March-July, leaf flushing peak period in the month of march where as flowering and fruiting activity peak period was found during the month of March—April and July—August respectively. So, this type of study will be helpful to give inferences in future whether the of climate change are giving pressure on the pe-

riodic behavior of medicinal plant species. Changed phenological behavior of plant species indicates the losing of plant diversity of the study region. Existing information on medicinal plants indicates habitat loss and has heightened they require for more proactive protection strategies. The need for adequate phenological information ahead of *ex-situ* conservation program is desirable. Low stocking density observed across the range and land use types is implicated for holistic conservation strategies and sustainability.

Keywords Phenology, Leaf fall, Leaf flush, Flowering, Fruiting.

Introduction

India is one of the mega biodiversity regions over the world. It also includes much plant diversity in different ecological areas followed by their environmental condition as well as adaptability of the plants in changeable climatic conditions. Plants are registered as efficient capability to regenerates as well as adjust in their variable localities. Plants diversity also supports the wealth of other species diversity in particular ecological areas and is remarkable for their major participation on environmental cleanup and supporting components for the presence and growth

Aadhan K*, Anand S. P.
 PG and Research Department of Botany, National College
 (Autonomous), Tiruchirappalli, Tamil Nadu, India
 e-mail : athanvivek@gmail.com
 *Corresponding author

of variable life forms. Plants are marked for their multifold utility in environment. On the basis of their importance for human beings are categorized as for food, fodder, fuel plants. Some plants also showing their efficiency to treat certain disorders these are referred as Medicinal and Aromatic plants (Patel 2014). India has well-off and diverse inheritance of biodiversity covering 10 bio-geographical zones. The Indian subcontinent is blessed with a wide variety of aromatic and medicinal plants. India natures enormous plant diversity and as many as 140 genera out of 5285 angiosperm species are endemic to the country (Botanical Survey of India 2001).

In nature it is often seen that each species has a specific period, month, season in a year during which its seeds germinate, seedlings grow or show maximum vegetative growth, leaves fall (if it is deciduous), flushing of new leaves, flowering and then fruiting. The study of all these periodic behavior of a species is called its phenology. In the life cycle of a plant each and every stage is greatly influenced by a number of environmental factors. The different stages of the plant species remain completely embedded in an environmental complex. It is very interesting to note that being fixed at a particular place, the requirement of germination, growth, flowering, fruiting, leaf fall of the species are met with at the same place but of course in different times of the year. There is a synchronization of phenological behavior of the species and the various factors of the surroundings that plants are spoken of biological clocks. This is mostly regulated by external signals from the environment. But the interactions of each and every species are different at different stages of their life cycle. Thus plant phenological study has great significance because it not only provides knowledge about the plant growth pattern but it also provides the idea on the effect of environment and selective pressure on flowering and fruiting behavior (Zhang et al. 2006).

Plants respond biologically to various parameters in the holocoenotic environment besides this, many extrinsic factors, e. g. the time of phenophase and seed dispersal are very important in distribution, survival and success in the establishment of a species in the community. Considering these facts, it is known that phenological studies are important for the conser-

vation of genetic resources and forest management as well as for a better understanding of ecological capabilities of plant species and community-level interactions. Another important attribute which decides the establishment of a species is population size. This is redulated by an array of environmental factors. In this study we have observed the times of phenophase and determined the influence of environmental variables on the population density of uncommon medicinal plants (Sundriyal et al. 1987, Duraisamy and Palsumy 2010).

Majority of them are reported to have medicinal properties. With increasing interest in herbal medicines worldwide, conservation of medicinal plants has assumed considerable importance. Conservation efforts including inventorying and documentation of the available medicinal plant diversity is the need of the hour. The study of phenology of medicinal plants in intact tropical ecosystems is also important if we are to learn more about the dynamics of medicinal plant species evolving in particular ecosystems, as well as how they may be excluded or successful in adjacent ecosystems; it is necessary to study first how species operate within the context of their respective environments. The study of gender distribution is useful in assessing the breeding systems in any ecosystem. There are many works concerning the breeding systems of tropical and sub-tropical forests of other regions of the world (Bawa 1974, Bullock 1985, Croat 1979, Flores and Schemske 1984, Freeman et al. 1980, Sivaraj and Krishnamurthy 1989, Sivaraj 1991).

Flowering and fruiting behaviors which are typical medicinal plants, sound decisions in sustainable *in-situ* and *ex-situ* conservation and management program for indigenous medicinal plants will require detailed knowledge of not only their taxonomy, natural regeneration pattern but also their current population pattern and their reproductive biology over the years, it is evident that substantial medicinal plants collections come from the wild without corresponding efforts at enrichment plantings or deliberate efforts to protect the remaining the germplasm because most collectors believe that there will always be enough in their natural habitat. However recent field observations indicate depletion in their gene pool (Oni

2001, 2004 and 2010). The most useful and main source of botanical information of a particular area is its floristic checklists. Floristic composition is a good floristic marker, because any kind of changing floristic compositions in different endogenous milieu show the existence of different ecological factors; leads to inter and intra-specific diversity (Safidkon et al. 2003). Floristic study of any given area helps to evaluate the plant wealth and its potential values. The local plants identification and introduction of an area is very important to introduce the specific species of local area and their occurrence, growing seasons, finding new species and also the effect of climatic conditions like over-grazing, drought and temperature on vegetation (Ali 2008, Ahmad et al. 2008).

Conservation of medicinal plants in natural ecosystem has assumed considerable importance. Conservation efforts including inventorying and documentation of the available medicinal plant diversity is the need of the hour. The learn of phenology of medicinal plants in intact tropical ecosystems is also important if we are to learn more about the dynamics of medicinal plant species evolving in particular ecosystems, as well as how they may be excluded or successful in adjacent ecosystems; it is necessary to study first how species operate within the context of their respective environments. The life history of plant species involves seed germination, vegetative growth, flowering, fruit formation, seed maturation, leaf fall, seed dispersal and death. Environmental factors influence the phenological events can be recorded diagrammatically month wise and season wise and provide valuable information (Barman et al. 2014).

Estimate that nearly 90% medicinal plants in use are collected the wild, in which 70% collection involves destructive harvesting. Habitat degradation due to prevailing biotic and non-biotic factors has caused to loss of various leaves of diversity and increasing risk of extinction to many high value species (Nayar and Sastry 1987, 1988 and 1990). Which need to understood to ascertain appropriate production site for any plant. Among important parameters, study of phenological behavior of any wild plants species, which is being targeted for cultivation is a pre-requisite exercise, it is helpful in developing and standardizing agrotechniques for targeted plant

species. Identification of phenological stages is incredibly critical (Sanz-Cortes et al. 2002). Phenology on the other hand is the study of growth buds, leaf flushing anthesis, fruiting and leaf fall in relation to season or years with climatic factors. It is the relationship of plant growth stage and calendar date. The calendar is based on the solar year. The information of phenology shows relationship of plant growth to seasonal changes and changes in length of day light or photoperiod to program their growth stage and biological activities appropriated with the seasonal condition (Manske 2006).

The flowering and fruiting could be correlated with climatic conditions for off spring's survival (Van Sachaik et al. 1993). The fruiting occurred at being of rainy season which helps survival of seeds to the exposure to predators and provides maximum time to seedlings for development of root system. It is now widely accepted that different biological interactions and phylogenetic relationships help in shaping the phenological patterns (Hamann 2004). Flowering time has been used as reproductive characters in taxonomic keys and in classification (Devis and Heywood 1973). Therefore plant phenology permits a calendar to construct the growth activity of plants especially the period of new leaf bud, appearance of mature leaves, flowers, bud, appearance of mature leaves, flowers, bud initiation, formation of mature flowers, young fruits formation and seeds maturity. Hence, it is very important to know about the exact timing of various phenological events for a particular tree or shrub species to raise the nursery in time and for the need of animal and human being for various purpose (Das and Pandey 2007, Sundriyal et al. 1987).

The life history of plant species involves seed germination, vegetative growth, flowering, fruit formation, seed maturation, leaf fall, seed dispersal and death. A study of the date and time of occurrence of these events is called phenology. Environmental factors influence the phenological events can be recorded diagrammatically month wise and season wise and provide valuable information such as diagram is called a phenogram. Study of phenology is important from the point of view of the conservation of tree genetics resources and forestry management as well as better understanding of the ecological adaptations. The study

of plant phenological provides knowledge about the pattern of plant growth and development as well as the effect of environment and selective pressures on flowering and fruiting behavior (Zhang et al. 2005). Review of literature indicates that our understanding on phenological behavior of medicinal plants under cultivation is very poor (Butola and Badola 2007, Sefidkon et al. 2003, Butola 2009, Vashistha et al. 2006). Considering the significance of phenological studies of plant species of a locality the present work was undertaken over a period 3 years (2015—2017). This was carried out to understand the response of some medicinal plant species to climatic factors and the periodicity of seasons of the Sadhuragiri hills.

Material and Methods

Study area

Sadhuagiri hills are situated in Southern Western Ghats comes under The Srivilliputhur Grizzled Squirrel Wildling sanctuary Srivilliputhur Taluk, Virudhunagar and Theni district. The elevation of Sadhuragiri is 1900 meters (3,937.0 ft.) msl in Western Ghats of South India. It lies between $9^{\circ}.42' - 9^{\circ}.44''$ West latitude and between $77^{\circ}.37' - 77^{\circ}.41''$ East longitude. Sadhuragiri is in an area with a Tropical evergreen forest, Semi evergreen forest and Mixed deciduous forest climate. The only tribal community residing in this region is Hindu Paliyar Tribes (Fig. 1).

Methods

The flowering fruiting periods and gender distribution and breeding systems of 165 medicinal plants occurring in the vegetation were recorded during the year monthly field trips to the Sadhuragiri hills, Western Ghats. They were examined in relation to fruit type (fleshy or dry) and vegetation type (scrub, deciduous, semi-evergreen) and habit form (Arborescent and Herbaceous). Arborescent taxa included shrubs, woody climbers, lianas and trees, while herbaceous taxa comprised herbs, slender twiners, climbers, epiphytes and parasites. Nomenclature for designating various categories of gender distribution has been followed from Richards (1986). For purposes of correlative analyses polygamodioecious species were considered Dioecious, similarly Polygamomonocious and Gynomonocious species were considered Monoecious. For purpose of computation the taxa that flowered and fruited throughout the year were listed as flowering and fruiting in all months. In addition, data obtained from the following sources were used : Herbarium specimens available at the Botanical Survey of India (BSI) and Rapinat Herbarium (RHT). Published floras (Jain 1963, Gamble 1935, Matthew 1991, Nair 1983, Henry 1987. Field books available at the Botanical Survey of India, Coimbatore, with phenological notes on 90 days, distributed all through the year. Field books available at the Rapinat Herbarium, Triuchirappalli, with phenological notes on 40 days, for collection made in this area during the years 2015—2017.

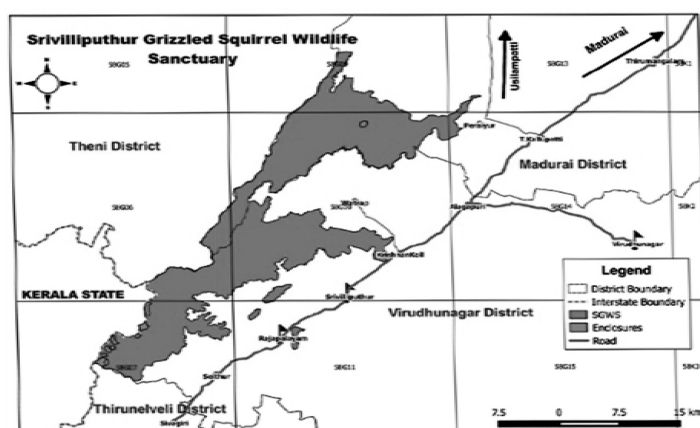


Fig. 1. The map over view of study region.

Table 1. List out the phenological study on the some medicinal plant species in Sadhuragiri hills. ALT and Vegetation type : Tropical thorn forest (300–500 m), Moist deciduous forest 500–700, Low evergreen forest (>800 m), Semievergreen forest >900, Grass land <1000, Growth form : Herbaceous–H, Arborscent–A breeding technology : Hermaphrodite –H, Monoecious–M, Dioecious–D, Polygamodioecious–Pd, Polygamomonocious–Pm, Gynomonocious–Gm, Fruit Type : DD–Dry dehiscent, DI–Dry indehiscent, F–Fleshy, F (I) – Fleshy (with one seeded), F (A) –Fleshy (with appendages).

Sl. No.	Name of the plant species	Family	AI (m)	VT	GF	BT	Flowering period
1.	<i>Abrus precatorius</i> L.	Fabaceae	500	TTE	H	H	Jan-Feb
2.	<i>Abutilon indicum</i> (L.) Sweet	Malvaceae	500	TTE	A	H	Jan-Apr
3.	<i>Abutilon polyandrum</i> (Roxb.) Wight & Arn.	Malvaceae	700	MDE	A	H	Jan-Apr
4.	<i>Acacia nilotica</i> (L.) Willd. Ex Del. Subsp. Indica.	Mimosaceae	500	TTE	A	H	Aug-Nov
5.	<i>Acacia planifrons</i> Wight & Arn.	Mimosaceae	700	MDF	H	H	Aug-Nov
6.	<i>Acacia auriculiformis</i> Benth.	Mimosaceae	700	MDF	A	H	Aug-Nov
7.	<i>Acalypha ciliata</i> Forrsk	Euphorbiaceae	800	LEF	H	M	Jan-Dec
8.	<i>Acalypha indica</i> L.	Euphorbiaceae	900	SEF	H	M	Jan-Dec
9.	<i>Acalypha racemosa</i> Wall. ex.Baill.	Euphorbiaceae	800	LEF	H	M	Jan-Dec
10.	<i>Achyranthes aspera</i> L.	Amaranthaceae	500	TTF	H	H	Jan-Dec
11.	<i>Achyranthes bidentata</i> Blume	Amaranthaceae	700	MDF	H	H	Jan-Dec
12.	<i>Adhatoda zeylanica</i> Medik.	Acanthaceae	600	MDF	H	H	Feb-Mar
13.	<i>Aegle marmelos</i> (L.) Correa ex. Schultz	Rutaceae	700	MDF	A	H	Mar-May
14.	<i>Aerva lanata</i> (L.) Juss.ex Schult.	Amaranthaceae	600	MDF	H	H	Jan-Mar
15.	<i>Agave americana</i> L.	Agavaceae	500	TTF	A	H	Apr-May
16.	<i>Ageratum conyzoides</i> L.	Asteraceae	700	MDF	H	H	Jan-Apr
17.	<i>Alangium salviifolium</i> (L.f.) Wangerin	Alangiaceae	800	LEF	H	H	Feb-Apr
18.	<i>Albizia lebbek</i> (L.) Benth	Mimosaceae	600	MDF	A	H	Feb-Apr
19.	<i>Alstonia scholaris</i> (L.) R. Br.	Apocynaceae	800	LEF	A	H	Nov-Dec
20.	<i>Alternanthera sessilis</i> (L.) R.Br. ex DC.	Amaranthaceae	500	TTF	H	H	Jan-Dec
21.	<i>Alysicarpus vaginalis</i> (L.) DC.	Fabaceae	500	TTF	H	H	Jan-Dec
22.	<i>Amaranthus spinosus</i> L.	Amaranthaceae	600	MDF	H	M	Jan-Dec
23.	<i>Amaranthus viridis</i> L.	Amaranthaceae	600	MDF	H	M	Jan-Dec
24.	<i>Anacardium occidentale</i> L.	Anacardiaceae	500	TTF	A	D	Feb-Mar
25.	<i>Andrographis paniculata</i> (Burm.f.) Wall.ex.Nees	Acanthaceae	700	MDF	H	H	May-Jul
26.	<i>Anisomeles malabarica</i> R.Br.ex.Sims.	Lamiaceae	500	TTF	H	H	Jan-Dec
27.	<i>Annona reticulata</i> L.	Annonaceae	800	LEF	A	D	Dec-Mar
28.	<i>Annona squamosa</i> L.	Annonaceae	700	MDF	A	D	Mar-May
29.	<i>Apama siliquosa</i> Lam.	Aristolochiaceae	800	LEF	A	H	Mar-May
30.	<i>Aristolochia bracteolata</i> Lam.	Aristolochiaceae	500	TTF	H	H	Feb-Apr
31.	<i>Aristolochia indica</i> L.	Aristolochiaceae	700	MDF	H	H	Feb-Apr
32.	<i>Asparagus racemosus</i> Willd.	Liliaceae	700	MDF	H	H	Oct-Nov
33.	<i>Atalantia monophylla</i> (L.) Corr. Serr.	Rutaceae	800	LEF	A	H	Jan-Apr
34.	<i>Averrhoa carambola</i> L.	Oxalidaceae	900	SEF	A	H	Feb-Mar
35.	<i>Bacopa monneria</i> L.Wettst.	Scrophulariaceae	800	LEF	H	H	Jun-Sep
36.	<i>Barleria buxifolia</i> L.	Acanthaceae	500	TTF	H	H	Jan-Mar
37.	<i>Bauhinia racemosa</i> LAM.	Caesalpiniaceae	800	LEF	A	H	Jan-Mar
38.	<i>Benkara malabarica</i> (LAM.)	Rubiaceae	600	MDF	A	H	Jan-Mar
39.	<i>Biophytum sensitivum</i> (L.) DC.	Oxalidaceae	1000	GL	H	H	Sep-Oct
40.	<i>Blepharis boerhaviifolia</i> Pers.	Acanthaceae	700	MDF	H	H	Jan-Feb
41.	<i>Boerhaavia diffusa</i> L.	Nyctaginaceae	500	TTF	H	H	Oct-Nov
42.	<i>Bombax pentandrum</i> JACQ.	Bombacaceae	600	MDF	A	H	Jan-Mar
43.	<i>Boswellia glabra</i> Roxb.	Burseraceae	700	MDF	A	H	Jan-Apr
44.	<i>Butea monosperma</i> (Lam.) Taub.	Fabaceae	900	SEF	A	H	Mar-Apr
45.	<i>Calophyllum inophyllum</i> L.	Clusiaceae	900	SEF	A	H	Mar-Apr
46.	<i>Calotropis gigantea</i> (L.) R.Br.	Asclepiadaceae	500	TTF	H	H	Feb-May
47.	<i>Cannabis sativa</i> L.	Cannabinaceae	700	MDF	H	H	Jan-Mar
48.	<i>Canthium coromandelicum</i> (N. Burm.)	Rubiaceae	600	MDF	A	H	Sep-Nov
49.	<i>Canthium parviflorum</i> Lam.	Rubiaceae	800	LEF	H	H	Sep-Nov
50.	<i>Capparis septaria</i> L.	Capparaceae	700	MDF	H	H	Mar-Apr
51.	<i>Capparis zeylanica</i> L.	Capparaceae	600	MDF	H	H	Mar-Apr

Table 1. Continued.

Sl. No.	Name of the plant species	Family	AI (m)	VT	GT	BT	Flowering period
52.	<i>Caralluma attemata</i> Wight	Asclepiadaceae	800	LEF	H	H	Apr-May
53.	<i>Caralluma umbellata</i> Haw.	Asclepiadaceae	700	MDF	H	H	Apr-May
54.	<i>Cardiospermum helicacabum</i> L.	Sapindaceae	500	TTF	H	H	Jan-Feb
55.	<i>Careya arborea</i> Roxb.	Lecythidaceae	800	LEF	A	H	Mar-Apr
56.	<i>Carissa carandas</i> L.	Apocynaceae	500	TTF	A	H	Feb-May
57.	<i>Carissa spinarum</i> L.	Apocynaceae	600	MDF	A	H	Feb-May
58.	<i>Caryota urens</i> L.	Palmaceae	900	SEF	A	H	Jan-Apr
59.	<i>Cassia auriculata</i> L.	Caesalpiniaceae	500	TTF	A	H	Jan-Mar
60.	<i>Cassia fistula</i> L.	Caesalpiniaceae	700	MDF	A	H	Jan-Mar
61.	<i>Cassia tora</i> L.	Caesalpiniaceae	700	LEF	H	H	Jan-Mar
62.	<i>Catharanthus roseus</i> (L.) G.Don	Apocynaceae	600	MDF	H	H	Jan-Dec
63.	<i>Celastrus paniculatus</i> Willd.	Celastraceae	800	LEF	H	PM	Jan-Mar
64.	<i>Centella asiatica</i> (L.) urban	Umbelliferae	600	MDF	H	H	Jan-Dec
65.	<i>Ceropegia candelabrum</i> L. Subsp.	Asclepiadaceae	900	SEF	H	H	Apr-May
66.	<i>Ceropegia juncea</i> Roxb.	Asclepiadaceae	1000	GL	H	H	Apr-May
67.	<i>Chenopodium ambrosioides</i> L.	Chenopodiaceae	900	SEF	H	H	Jan-Dec
68.	<i>Cissus quadrangularis</i> L.	Vitaceae	500	TTF	H	H	Feb-Mar
69.	<i>Clitoria ternatea</i> L.	Fabaceae	500	TTF	H	H	Jan-Dec
70.	<i>Coccinia grandis</i> (L.) Voigt.	Cucurbitaceae	500	TTF	H	H	Jan-Dec
71.	<i>Cocculus laurifolius</i> DC.	Menispermaceae	700	MDF	H	D	Feb-Mar
72.	<i>Cyanthillium albicans</i> (DC.) H. Rob.	Asteraceae	700	MDF	H	H	Feb-May
73.	<i>Cynodon dactylon</i> (L.) Pers.	Poaceae	700	MDF	H	H	Jan-Dec
74.	<i>Dalbergia latifolia</i> Roxb.	Fabaceae	800	LEF	A	H	Jul-Aug
75.	<i>Datura metel</i> L.	Solanaceae	700	MDF	H	H	Jan-Dec
76.	<i>Decalapis hamiltonii</i> Wight & Arn.	Asclepiadaceae	900	SEF	A	H	Feb-Mar
77.	<i>Desmodium triflorum</i> (L.) DC.	Fabaceae	800	LEF	H	H	Jan-Feb
78.	<i>Dillenia indica</i> L.	Dilleniaceae	900	SEF	A	H	Mar-Apr
79.	<i>Dioscorea pentaphylla</i> L.	Dioscoreaceae	800	LEF	H	M	Feb-Mar
80.	<i>Diospyros ebenum</i> J.Koenig ex.Retz.	Ebenaceae	800	LEF	A	M	Mar-Apr
81.	<i>Diospyros melanoxylon</i> Roxb.	Ebenaceae	800	LEF	H	M	Mar-Apr
82.	<i>Diospyros montana</i> Roxb.	Ebenaceae	800	LEF	A	M	Mar-Apr
83.	<i>Diospyros paniculata</i> Dalz	Ebenaceae	700	MDF	A	M	Mar-Apr
84.	<i>Eclipta alba</i> (L.) Massk.	Asteraceae	600	MDF	H	GM	Jan-Feb
85.	<i>Elaeocarpus recurvatus</i> Corner	Elaeocarpaceae	900	SEF	A	H	Apr-May
86.	<i>Elaeocarpus tuberculatus</i> Roxb.	Elaeocarpaceae	800	LEF	H	H	Apr-May
87.	<i>Emblica officinalis</i> Gaertner	Euphorbiaceae	900	SEF	A	H	Mar-Apr
88.	<i>Erythrina indica</i> L.	Fabaceae	800	LEF	A	H	Jan-Mar
89.	<i>Euphorbia hirta</i> L.	Euphorbiaceae	500	TTF	H	M	Jan-Dec
90.	<i>Evolvulus alsinoides</i> (L.) L.	Convolvulaceae	600	MDF	H	H	Jan-Dec
91.	<i>Feronia elephantum</i> Corr.serr.	Rutaceae	800	LEF	A	H	Jan-Mar
92.	<i>Ficus bendhalensis</i> L.	Moraceae	700	MDF	A	PM	Apr-May
93.	<i>Givotia moluccana</i> (L.) Sreem.	Euphorbiaceae	700	MDF	A	H	Jan-Mar
94.	<i>Gloriosa superba</i> L.	Liliaceae	800	LEF	H	H	Jan-Feb
95.	<i>Gmelina arborrea</i> Roxb.	Verbenaceae	600	MDF	A	H	Jan-Mar
96.	<i>Gmelina asiatica</i> L.	Verbenaceae	800	LEF	H	H	Jan-Feb
97.	<i>Grewia flavescens</i> Juss	Tiliaceae	700	LEF	A	H	Mar-Apr
98.	<i>Grewia hirsuta</i> vahl	Tiliaceae	600	MDF	A	H	Mar-Apr
99.	<i>Grewia orientalis</i> L.	Tiliaceae	800	LEF	A	H	Mar-Apr
100.	<i>Grewia tiliifolia</i> Vahl	Tiliaceae	700	LEF	A	H	Mar-Apr
101.	<i>Gymnema sylvestre</i> (Retz.) R.Br. ex Schult	Asclepiadaceae	700	MDF	H	H	Jan-Mar
102.	<i>Helicteres isora</i> L.	Sterculiaceae	800	LEF	A	H	Sep-Dec
103.	<i>Hemidesmus indicus</i> (L.) R. Br.	Asclepiadaceae	800	LEF	H	H	Jan-Mar
104.	<i>Hibiscus vitifolius</i> L.	Malvaceae	500	TTF	H	H	Jan-Feb
105.	<i>Hibiscus canescens</i> B. Heyne ex Wall.	Malvaceae	600	MDF	H	H	Jan-Feb
106.	<i>Hibiscus ovalifolius</i> (Forssk.) Vahl	Malvaceae	600	MDF	H	H	Jan-Feb

Table 1. Continued.

Sl. No.	Name of the plant species	Family	AI (m)	VT	GT	BT	Flowering period
107.	<i>Hibiscus planifolius</i> Sweet	Malvaceae	800	LEF	H	H	Jan-Feb
108.	<i>Hybanthus enneasper mus</i> (L.) F. Muell.	Violaceae	700	MDF	H	H	Mar-Apr
109.	<i>Hydnocarpus macrocapra</i> (Bedd.)Warp	Flacourtiaceae	800	LEF	A	H	Feb-Apr
110.	<i>Hyptis suaveolens</i> (L.) Poit	Lamiaceae	500	TTF	H	H	Jan-Feb
111.	<i>Ichnocarpus frutescens</i> (L.)W.T.Aiton	Apocynaceae	700	MDF	H	H	Jan-Mar
112.	<i>Jatropha curcas</i> L.	Euphorbiaceae	800	LEF	A	M	Jan-Mar
113.	<i>Justicia gendarussa</i> Burm.f.	Acanthaceae	500	TTF	H	H	Jan-Mar
114.	<i>Leonotis nepetifolia</i> (L.) R. Br.	Lamiaceae	900	SEF	H	H	Jan-Feb
115.	<i>Leucas lanceifolia</i> Desf.	Lamiaceae	900	SEF	H	H	Jan-Feb
116.	<i>Leucas biflora</i> (Vahl) Sm.	Lamiaceae	900	SEF	H	H	Jan-Feb
117.	<i>Lindernia parviflora</i> (Roxb.) Haines	Scrophulariaceae	600	MDF	H	H	Jan-Feb
118.	<i>Mallotus philippensis</i> Muell. Arg.	Euphorbiaceae	900	SEF	A	D	Jan-Dec
119.	<i>Melia azedarach</i> L.	Meliaceae	700	MDF	A	H	Mar-Apr
120.	<i>Michelia champaca</i> L.	Magnoliaceae	800	LEF	A	H	Mar-Apr
121.	<i>Miliusa eriocarpa</i> Dunn	Annonaceae	900	SEF	A	H	Jan-Mar
122.	<i>Mimosa pudica</i> L.	Mimosaceae	800	LEF	H	PM	Jan-Feb
123.	<i>Mucuna prurita</i> Hook.	Fabaceae	900	SEF	H	H	Mar-Apr
124.	<i>Murraya koenigii</i> (L.) Spreng.	Rutaceae	700	MDF	A	H	Jan-Mar
125.	<i>Murraya paniculata</i> (L.) Jack	Rutaceae	800	LEF	A	H	Jan-Mar
126.	<i>Ocimum canum</i> Sims.	Lamiaceae	500	TTF	H	H	Feb-Mar
127.	<i>Oldenlandia corymbosa</i> L.	Rubiaceae	500	TTF	H	H	Feb-Mar
128.	<i>Orthosiphon thymiflorus</i> (Roth) Sleesen	Lamiaceae	900	SEF	H	H	Jan-Mar
129.	<i>Oxalis corniculata</i> L.	Oxalidaceae	900	SEF	H	H	Jan-Mar
130.	<i>Passiflora foetida</i> L.	Passifloraceae	700	MDF	H	H	Mar-Apr
131.	<i>Pavetta indica</i> L.	Rubiaceae	600	MDF	A	H	Jan-Feb
132.	<i>Phyllanthus virgatus</i> G. Forst.	Euphorbiaceae	800	LEF	H	M	Jan-Feb
133.	<i>Pithecolobium dulce</i> (Roxb.) Benth	Mimosaceae	800	LEF	A	PM	Jan-Feb
134.	<i>Plectranthus coleoides</i> Benth.	Lamiaceae	700	MDF	H	H	Mar-Apr
135.	<i>Plumbago zeylanica</i> L.	Plumbaginaceae	800	LEF	H	H	Feb-Mar
136.	<i>Polygala arvensis</i> Willd.	Polygalaceae	800	LEF	H	H	Jan-Feb
137.	<i>Portulaca oleracea</i> L.	Portulacaceae	500	TTF	H	H	Feb-Mar
138.	<i>Premna tomentosa</i> Willd.	Verbenaceae	900	SEF	A	H	Feb-Mar
139.	<i>Psilanthus wightianus</i> (Wall.ex Wight & Arn.) J. F. Leroy	Rubiaceae	700	MDF	H	H	Feb-Mar
140.	<i>Pterocarpus marsupium</i> Roxb.	Fabaceae	900	SEF	A	H	Sep-Oct
141.	<i>Ricinus communis</i> L.	Euphorbiaceae	500	TTF	A	H	Jan-Feb
142.	<i>Rubia cordifolia</i> L.	Rubiaceae	600	LEF	H	H	Mar-Apr
143.	<i>Smilax wightii</i> A. DC.	Smilacaceae	800	LEF	H	H	Mar-Apr
144.	<i>Solanum nigrum</i> L.	Solanaceae	500	TTF	H	H	Feb-Mar
145.	<i>Solanum pubescens</i> Ruiz & Pav.	Solanaceae	600	MDF	H	H	Feb-Mar
146.	<i>Syzygium cuminii</i> (L.) Skeels.	Myrtaceae	800	LEF	A	H	Jun-Jul
147.	<i>Tephrosia purpurea</i> (L.) Pers.	Fabaceae	500	TTF	H	H	Jan-Feb
148.	<i>Terminalia bellirica</i> (Gaertn.) Roxb.	Combretaceae	900	SEF	A	PM	Mar-May
149.	<i>Terminalia arjuna</i> (Roxb. ex DC.) Wight & Arn.	Combretaceae	900	SEF	A	PM	Mar-Apr
150.	<i>Terminalia chebula</i> Retz.	Combretaceae	900	SEF	A	PM	Mar-Apr
151.	<i>Tinospora cordifolia</i> (Thunb.) Miers.	Menispermaceae	600	MDF	H	D	Jan-Feb
152.	<i>Trema orientalis</i> (L.) Blume	Ulmaceae	900	SEF	A	D	Mar-Apr
153.	<i>Tribulus terrestris</i> L.	Zygophyllaceae	500	TTF	H	H	Jan-Feb
154.	<i>Trichodesma zeylanicum</i> (Burm.f.)	Boraginaceae	700	MDF	H	H	Jan-Feb
155.	<i>Vanda tessellata</i> (Roxb.) Don.	Orchidaceae	700	MDF	H	H	Jan-Feb
156.	<i>Ventilago maderasptana</i> Gaertner	Rhamnaceae	500	TTF	H	H	Feb-Mar
157.	<i>Viscum angulatum</i> B. Heyne ex DC.	Viscaceae	800	LEF	H	H	Jan-Feb
158.	<i>Vitex negundo</i> L.	Verbenaceae	500	TTF	A	H	Feb-Mar
159.	<i>Walsure trifoliolata</i> (A. Juss.) Harms	Meliaceae	900	SEF	A	H	Jan-Feb
160.	<i>Waltheria indica</i> L.	Sterculiaceae	900	SEF	A	H	Feb-Mar

Table 1. Continued.

Sl. No.	Name of the plant species	Family	AI (m)	VT	GT	Bt	Flowering period
161.	<i>Withania somniferum</i> (L.) Dunal	Solanaceae	700	MDF	H	H	Jan-Feb
162.	<i>Wrightia tinctoria</i> (Roxb.) R. Br.	Apocynaceae	800	LEF	A	H	Mar-Apr
163.	<i>Xanthium indicum</i> KOEN.	Asteraceae	700	MDF	H	M	Jan-Feb
164.	<i>Ziziphus oenoplia</i> (L.) Miller	Rhamnaceae	500	TTF	A	H	Feb-Mar
165.	<i>Zizyphus nummularia</i> (Burm. F.) Wights	Rhamnaceae	700	MDF	A	H	Feb-Mar

Table 1. Continued.

Sl. No.	Fruiting period	FT	Leaf fall period	Leaf flush period	Conservative part	Reproductive cost	
						Seed setting	Seed germinative
1.	Mar-Apr	DD	May-Jun	Oct-Nov	Seed	Minimum	Poor
2.	Mar-May	DD	Jun-Aug	Oct-Dec	Seed	Maximum	Very good
3.	Mar-May	DD	Jun-Aug	Oct-Dec	Seed	Maximum	Very good
4.	Sep-Nov	DD	Mar-May	Jan-Dec	Seed	Minimum	Good
5.	Sep-Nov	DD	Apr-Jun	Jan-Dec	Seed	Minimum	Good
6.	Sep-Nov	DD	Mar-May	Jan-Dec	Seed	Minimum	Good
7.	Jan-Dec	D	Jan-Dec	Jan-Dec	Seed	Few	Good
8.	Jan-Dec	D	Jan-Dec	Jan-Dec	Seed	Few	Good
9.	Jan-Dec	D	Jan-Dec	Jan-Dec	Seed	Few	Good
10.	Jan-Dec	DI	Jan-Dec	Jan-Dec	Seed	Maximum	Very good
11.	Jan-Dec	DI	Jan-Dec	Jan-Dec	Seed	Minimum	Very good
12.	Apr-May	D	Jun-Aug	Aug-Sep	Seed	Few	Good
13.	May-Jun	F (A)	Dec-Jan	Dec-Jan	Seed	Minimum	Poor
14.	Jan-Apr	DD	Dec-Jan	Dec-Jan	Rhizome	Maximum	Good
15.	May-Jun	DD			Stem/Seed	Few	Good
16.	Jan-May	DI	Jan-Dec	Jan-Dec	Seed	Maximum	Very good
17.	Mar-Jun	F (I)	Dec-Jan	Mar-Apr	Seed	Average	Very good
18.	Mar-Jun	DD	Dec-Jan	Feb-Apr	Seed	Minimum	Good
19.	Jan-Mar	DD	Feb-Apr	May-Jul	Stem	Minimum	Good
20.	Jan-Dec	DI	Jan-Dec	Jan-Dec	Seed	Maximum	Very good
21.	Jan-Dec	DI	Jan-Dec	Jan-Dec	Seed	Minimum	Good
22.	Jan-Dec	DI	Jan-Dec	Jan-Dec	Seed	Maximum	Very good
23.	Jan-Dec	DI	Jan-Dec	Jan-Dec	Seed	Maximum	Very good
24.	Apr-May	FI	Sep-Oct	Nov-Dec	Seed	Minimum	Good
25.	Aug-Sep	DD	Jan-Feb	Mar-Apr	Seed	Average	Good
26.	Jan-Dec	DI	Jan-Dec	Jan-Dec	Seed	Maximum	Good
27.	Jan-May	DD	Mar-Jun	Nov-Dec	Seed	minimum	Good
28.	Aug-Nov	F (A)	Jan-Feb	Mar-Apr	Seed	Minimum	Good
29.	Aug-Nov	F (A)	Jan-Feb	Mar-Apr	Seed	Average	Poor
30.	Mar-May	DI	Jan-Feb	Apr-May	Seed	Minimum	Poor
31.	Mar-May	DI	Jan-Feb	Apr-May	Seed	Minimum	Poor
32.	Sep-Jan	DD	Jun-Aug	Jun-Oct	Tuber	Few	Very good
33.	Feb-May	DI	Jan-May	Apr-May	Seed	Minimum	Good
34.	Mar-Apr	DI	Feb-Mar	Mar-Apr	Seed	Minimum	Good
35.	Oct-Dec	DD	Jun-Jul	Aug-Sep	seed	Minimum	Good
36.	Feb-May	DI	Sep-Jan	May-jun	Seed	Minimum	Good
37.	Feb-Jun	F (I)	Sep-Jan	May-Jun	Seed	Minimum	Good
38.	Feb-Jun	DI	Sep-Jan	May-Jun	Seed	Maximum	Good
39.	Nov-Dec	DD	Jun-Jul	Aug-Sep	Seed	Few	Poor
40.	Feb-Mar	DD	Sep-Oct	Nov-Dec	Seed	Minimum	Good
41.	Jan-Apr	DI	Aug-Sep	Oct-Dec	Seed	Minimum	Good
42.	Mar-Jun	DI	Sep-Oct	Nov-Dec	Seed	Minimum	Good
43.	Mar-Jul	F (I)	Aug-Sep	Oct-Jan	Seed	Few	Very poor

Table 1. Continued.

Sl. No.	Fruiting period	FT	Leaf fall period	Leaf flush period	Conservative part	Reproductive cost	
						Seed setting	Seed germinative
44.	Apr-Jun	F (I)	Jan-Feb	Feb-Mar	Seed	Minimum	Poor
45.	May-Apr	DD	Jan-Dec	Jan-Dec	Seed	Minimum	Good
46.	May-Aug	DD	Nov-Dec	Jan-Feb	Seed	Minimum	Good
47.	May-Jul	DI	Oct-Nov	Nov-Jan	Seed	Few	Poor
48.	Jan-Apr	F (I)	Apr-Jun	Apr-Jun	Seed	Few	Good
49.	Jan-Apr	F (I)	Dec-Mar	Apr-Jun	Seed	Minimum	Good
50.	Apr-Jul	F (A)	Jan-Mar	Mar-Aug	Seed	Minimum	Good
51.	Apr-Jul	F (A)	Jan-Mar	Mar-Aug	Seed	Few	Poor
52.	May-Jun	DD	Jan-Mar	Feb-May	Bulb	Few	Very poor
53.	May-Aug	DD	Jan-Mar	Feb-May	Bulb	Few	Very poor
54.	Mar-Apr	DI	Jul-Aug	Aug-Sep	Seed	Minimum	Good
55.	Apr-Jun	F	Jan-Feb	Mar-Apr	Seed	Few	Poor
56.	Mar-Jun	F	Jan-Mar	Mar-May	Seed	Minimum	Good
57.	Mar-Jun	F	Jan-Mar	Mar-May	Seed	Few	Poor
58.	Feb-Jun	F (I)	Apr-May	Jul-Sep	Seed	Few	Poor
59.	Feb-May	DI	Oct-Nov	Nov-Dec	Seed	Maximum	Good
60.	Feb-May	DI	Oct-Nov	Nov-Dec	Seed	Minimum	Good
61.	Feb-May	DI	Oct-Nov	Nov-Dec	Seed	Minimum	Good
62.	Jan-Dec	DD	Jan-Dec	Jan-Dec	Seed	Maximum	Very good
63.	Mar-May	F (A)	Sep-Oct	Oct-Nov	Seed	Minimum	Good
64.	Jan-Dec	DI			Seed	Maximum	Very good
65.	Jun-Aug	DD	Sep-Oct	Jan-Feb	Tuber	Few	Very poor
66.	Jun-Aug	DD	Sep-Oct	Jan-Feb	Tuber	Few	Very poor
67.	Jan-Dec	DI	Sep-Oct	Jan-Feb	Seed	Average	Good
68.	Apr-May	F	Sep-Nov	Jun-Jul	Stem	Minimum	Good
69.	Jan-Dec	DI	Sep-Oct	Jan-Feb	Seed	Minimum	Good
70.	Jan-Dec	F (A)	Sep-Oct	Jan-Feb	Seed	Maximum	Good
71.	Apr-May	F	Sep-Nov	Jun-Jul	Seed	Maximum	Good
72.	Mar-Jun	F (I)	Jan-Mar	Mar-May	Seed	Minimum	Good
73.	Jan-Dec	DI	May-Jul	Jun-Sep	Seed/Rhizome	Maximum	Very good
74.	Aug-Oct	DD	May-Jul	Jun-Sep	Seed	Few	Poor
75.	Jan-Dec	DD	May-Jul	Jun-Sep	Seed	Minimum	Good
76.	Apr-May	DD	Mar-Apr	Apr-Jun	Seed	Few	Poor
77.	Feb-Apr	DD	Nov-Dec	Apr-May	Seed	Minimum	Good
78.	Apr-Jun	DI	Jan-Mar	May-Jun	Seed/Stem	Minimum	Poor
79.	Mar-May	DD	Jan-Feb	Mar-May	Bulb	Few	Poor
80.	Apr-Jun	F	Jan-Mar	Feb-Apr	Seed	Few	Poor
81.	Apr-Jun	F	Jan-Mar	Feb-Apr	Seed	Few	Poor
82.	Apr-Jun	F	Jan-Mar	Feb-Apr	Seed	Few	Very poor
83.	Apr-Jun	F	Jan-Mar	Feb-Apr	Seed	Few	Poor
84.	Mar-Apr	DI	Oct-Nov	Nov-Dec	Seed	Maximum	Very good
85.	May-Jul	F (I)	Jan-Mar	Feb-Apr	Seed	Few	Nil
86.	May-Jul	F (I)	Jan-Mar	Feb-Apr	Seed	Few	Nil
87.	Apr-Jun	F (I)	Jan-Feb	Feb-Mar	Seed	Minimum	Good
88.	Mar-Apr	DD	Aug-Sep	Nov-Dec	Seed/Stem	Average	Good
89.	Jan-Dec	F (A)	Jan-Dec	Jan-Dec	Seed	Minimum	Very good
90.	Jan-Dec	DD	Jan-Dec	Jan-Dec	Seed	Minimum	Very Good
91.	Apr-May	F	Nov-Dec	Feb-Mar	Seed	Minimum	Good
92.	May-Jul	DI	Jan-Mar	Mar-Apr	Stem/Seed	Maximum	Very poor
93.	May-Jul	DI	Oct-Dec	Mar-Apr	Seed/Stem	Minimum	Poor
94.	Mar-Apr	DD	Sep-Oct	Nov-Dec	Seed	Minimum	Good
95.	Mar-Apr	DD	Sep-Oct	Nov-Dec	Seed	Minimum	Good
96.	Mar-Apr	DD	Apr-May	Jun-Jul	Seed	Minimum	Good
97.	May-Jun	F (I)	Jul-Aug	Sep-Oct	Seed	Minimum	Poor
98.	May-Jun	F (I)	Jul-Aug	Sep-Oct	Seed	Minimum	Poor

Table 1. Continued.

Sl. No.	Fruiting period	FT	Leaf fall period	Leaf flush period	Conservative part	Reproductive cost	
						Seed setting	Seed germinative
99.	May-Jun	F (I)	Jul-Aug	Sep-Oct	Seed	Minimum	Poor
100.	May-Jun	F (I)	Jul-Aug	Sep-Oct	Seed	Minimum	Poor
101.	Mar-Apr	DD	Sep-Oct	Nov-Dec	Seed	Few	Poor
102.	Dec-Mar	DD	Jan-Apr	Jun-Aug	Seed	Few	Nil
103.	Mar-Apr	DD	Sep-Oct	Nov-Dec	Root/Stem	Few	Very poor
104.	Mar-Apr	DD	Jun-Aug	Oct-Dec	Seed	Minimum	Good
105.	Mar-Apr	DD	Jun-Aug	Oct-Dec	Seed	Minimum	Good
106.	Mar-Apr	DD	Jun-Aug	Oct-Dec	Seed	Minimum	Good
107.	Mar-Apr	DD	Jun-Aug	Oct-Dec	Seed	Minimum	Good
108.	Apr-May	DI	Aug-Sep	Oct-Nov	Seed	Few	Poor
109.	Apr-May	F (I)	Jun-Jul	Aug-Sep	Seed	Few	Poor
110.	Mar-Apr	DD	May-Jun	Jun-Aug	Seed	Maximum	Very good
111.	Mar-Apr	DD	Jun-Jul	Aug-Sep	Seed	Few	Very poor
112.	Mar-Apr	DI	Jun-Jul	Aug-Sep	Seed/Stem	Minimum	Very good
113.	Mar-Apr	DI	Jun-Jul	Aug-Sep	Seed	Minimum	Good
114.	Mar-Apr	DD	Jun-Aug	Oct-Dec	Seed	Minimum	Poor
115.	Mar-Apr	DD	Jul-Sep	Aug-Nov	Seed	Minimum	Good
116.	Mar-Apr	DD	Jul-Sep	Aug-Nov	Seed	Minimum	Good
117.	Mar-Apr	DD	Jul-Sep	Aug-Nov	Seed	Minimum	Good
118.	Jan-Dec	DI	Jan-Dec	Jan-Dec	Seed/Stem	Minimum	Good
119.	Apr-May	F (I)	Jun-Jul	Aug-Sep	Seed/Stem	Maximum	Very good
120.	Apr-May	DD	Jun-Jul	Aug-Sep	Seed/Stem	Few	Poor
121.	Mar-May	F (I)	May-Aug	Aug-Oct	Seed	Few	Poor
122.	Feb-Apr	DD	May-Aug	Aug-Oct	Seed	Maximum	Very good
123.	Apr-May	DI	Jun-Jul	Aug-Sep	Seed	Few	Good
124.	Mar-Apr	F (I)	Jun-Jul	Aug-Sep	Seed	Minimum	Good
125.	Mar-Apr	F (I)	Jun-Jul	Aug-Sep	Seed	Minimum	Good
126.	Mar-Apr	DI	Jul-Sep	Aug-Nov	Seed	Maximum	Very good
127.	Mar-Apr	DD	Jul-Sep	Aug-Nov	Seed	Minimum	Very good
128.	Mar-Apr	DD	Jul-Sep	Aug-Nov	Seed	Minimum	Very good
129.	Mar-Apr	F (A)	May-Aug	Aug-Oct	Seed	Minimum	Good
130.	Apr-May	F (A)	Jun-Jul	Aug-Sep	Seed	Minimum	Good
131.	Mar-Apr	F (I)	Jul-Sep	Aug-Nov	Seed	Maximum	Good
132.	Mar-Apr	DD	Jul-Sep	Aug-Nov	Seed	Minimum	Good
133.	Mar-Apr	F (I)	Jul-Sep	Aug-Nov	Seed	Maximum	Good
134.	Apr-May	DD	Jun-Jul	Aug-Sep	Seed	Minimum	Good
135.	Mar-Apr	DI	Jul-Sep	Oct-Nov	Seed	Average	Poor
136.	Mar-Apr	DI	Jun-Aug	Oct-Dec	Seed	Few	Good
137.	Mar-May	DD	May-Jun	Jul-Aug	Seed/Stem	Minimum	Good
138.	Mar-May	DD	May-Jun	Jul-Aug	Seed	Minimum	Poor
139.	Mar-Apr	F (A)	Jul-Sep	Aug-Nov	Seed	Minimum	Poor
140.	Oct-Nov	F	Jan-Feb	Mar-Apr	Seed	Minimum	Poor
141.	Mar-Apr	DI	Jun-Aug	Oct-Dec	Seed	Maximum	Very good
142.	Mar-May	DD	May-Jun	Jul-Aug	Seed	Minimum	Good
143.	Apr-Jun	DI	Aug-Sep	Oct-Nov	Seed	Minimum	Poor
144.	Mar-Apr	F	Jul-Sep	Aug-Nov	Seed	Maximum	Very good
145.	Mar-Apr	F	Jul-Sep	Aug-Nov	Seed	Minimum	Good
146.	Jul-Aug	F	Mar-Apr	Apr-May	Seed	Minimum	Good
147.	Mar-Apr	DI	Jun-Aug	Oct-Dec	Seed	Maximum	Very good
148.	Apr-May	DD	Jun-Jul	Aug-Sep	Seed	Minimum	Poor
149.	Apr-May	DD	Jun-Jul	Aug-Sep	Seed	Minimum	Poor
150.	Apr-May	F (I)	Jun-Jul	Aug-Sep	Seed	Few	Very good
151.	Mar-Apr	F (A)	Jul-Sep	Aug-Nov	Seed	Average	Poor
152.	Apr-May	F (I)	Jun-Jul	Aug-Sep	Seed	Few	Poor
153.	Mar-May	D	Aug-Nov	Jul-Sep	Seed	Maximum	Very good

Table 1. Continued.

Sl. No.	Fruiting period	FT	Leaf fall period	Leaf flush period	Conservative part	Reproductive cost	Seed setting	Seed germinative
154.	Mar-Apr	F (I)	Jun-Aug	Oct-Dec	Seed	Minimum	Very good	
155.	Mar-Apr	DD	Jun-Aug	Oct-Dec	Bud	Few	Poor	
156.	Mar-Apr	F (I)	Jul-Sep	Aug-Nov	Seed	Minimum	Poor	
157.	Mar-Apr	DD	Jun-Aug	Oct-Dec	Stem	Few	Poor	
158.	Mar-Apr	DD	Jul-Sep	Aug-Nov	Seed/Stem	Minimum	Good	
159.	Mar-Apr	DI	Jun-Aug	Oct-Dec	Seed	Minimum	Good	
160.	Apr-May	DD	Jul-Sep	Aug-Nov	Seed	Few	Poor	
161.	Mar-Apr	F	Jul-Aug	Aug-Sep	Seed	Minimum	Good	
162.	Apr-May	DI	Jun-Jul	Jul-Aug	Seed/Stem	Minimum	Good	
163.	Mar-Apr	DI	Jun-Aug	Oct-Dec	Seed	Average	Poor	
164.	Apr-May	F (I)	Jul-Sep	Aug-Nov	Seed	Minimum	Good	
165.	Apr-May	F (I)	Jul-Sep	Aug-Nov	Seed	Minimum	Good	

Results and Discussion

The study of floristic ecology and phenology of medicinal plants in intact tropical ecosystems is important to understand the dynamics of medicinal plant species evolving in particular ecosystems. The Southern most hill of the Western Ghats, Sadhuragiri comprises 165 taxa in distribution as Moist deciduous forest (34.5%), Semi evergreen forest (26.06%), Tropical thorn forest (18.18%), Low evergreen forest (16.36%) and Grass land (1.21%) vegetation types. Among the gender distribution, hermaphrodite category dominated (82%) in the medicinal flora followed by monoecy (8.4%) and dioecy (4.2%) (Table 1 and Fig.2).

Popular medicinal arborescent taxa was found to flower during the months of March and April (dry period) and only a few are in bloom during August

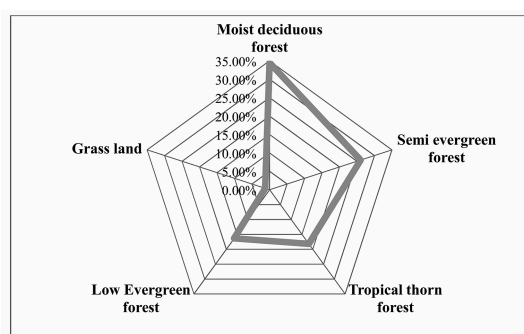


Fig. 2. Distribution of phenology study medicinal plants accumulated different forest types.

and December, while herbaceous taxa flowered during December-February. Majority of tree taxa were found in fruit during the dry season (April-May) with a peak during April while the herbaceous medicinal plants fruited during March-June (Fig. 3). The nature of fruits types recorded was Dry dehiscent (66 sps), Dry indehiscent (43 sps), Fleshy (with one seeded) (29 sps), Fleshy (15 sps), Fleshy (with appendages) (12 sps). Higher temperature conditions were observed to be conducive to flowering and fruiting among the arborescent taxa, whereas cooler conditions were favorable for herbaceous taxa. In Sadhuragiri hills, the fruiting phenology pattern closely follows that of the flowering.

The majority of tree taxa fruits for the period of the dry season with a peak during April-May while the peak fruiting period recorded for herbaceous taxa was during January-February. The time interval between flowering and fruiting was normally 30–55

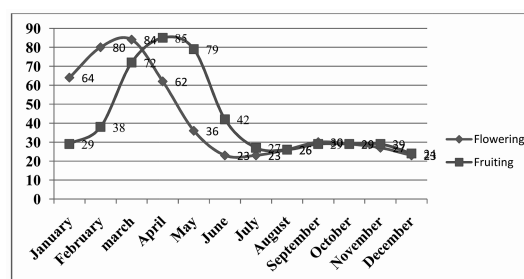


Fig. 3. Flowering and Fruiting pattern in some medicinal plants of Sadhuragiri hills.

days. Three species viz., *Buddleja asiatica* (Buddlejaceae), *Cissus quadrangularis* (Vitaceae) and *Ricinus communis* (Euphorbiaceae) were observed to flower and fruit throughout the year while 5 species namely *Mallotus philippensis* (Euphorbiaceae), *Michelia champaca* (Magnoliaceae), *Tithonia diversifolia* (Asteraceae), *Trema orientalis* (Ulmaceae) and *Solanum indicum* (Solanaceae) flowered and fruited in 2 spells (Sivaraj et al. 2014). The pattern of leaf fall and leaf flush recorded in the study region highest leaf fall occurred in March-July while the least was observed during June-October. The maximum leaf flush was seen in July-August while the least was recorded in November-January (Fig.4).

Phenology is the art of observing life cycle or activities of plants in their temporal occurrence throughout the growing season. Phenological and phenomenological variations of the plants are the product of interaction between genotype and environment. However, these modifications in plants may be reversible when plants are grown under diverse climatic conditions (Bhatt and Purohit 1984). The data analysis it was found that leaf fall initiation was a periodic activity for most of the species. In popular of the species leaf fall starts in the month of November and December with a peak in the last part of January (30%) to first part of February (50 %). Later than shedding of older leaves new leaf initiation starts in the species, the time period of this activity seen to be different in different species. But it can be said that new leaf formation started in majority of species in the month of February (25%) continued up to May

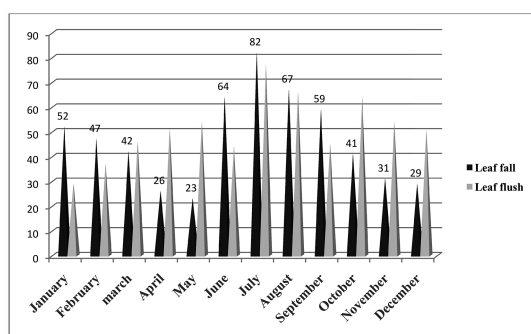


Fig. 4. Leaf all and leaf flush in some medicinal plants of Sadhuragiri hills.

(30%) with a peak in the month of March (50%) that is before the outset of monsoon. Among 40 species, 75% showed brief leaf flushing activity whereas only about 25% exhibited extended leaf activation (Barman et al. 2014).

In the majority cases leaf flushing was seasonal and occurred simply once in year. Under this category, leaf flushing occurred in the majority of the species towards the end of the dry season. Nine species alone, namely *Garuga pinnata* (Burseraceae), *Pterocarpus marsupium* (Papilionaceae), *Cinnamomum camphorum* (Lauraceae), *Strychnos nux-vomica* (Loganiaceae), *Feronia elephantum* (Rutaceae), *Homonoia riparia* (Euphorbiaceae), *Atalantia monophylla* (Rutaceae), *Citrus medica* (Rutaceae) and *Aglaiia roxburghiana* (Meliaceae) produced new leaves during the cooler months of December and January. The growth of new leaves, when it was irregular occurred at periodic or intermittent intervals. In *Cinnamomum camphora* (Lauraceae) *Boswellia glabra* (Burseraceae) and *Pterocarpus marsupium* (Papilionaceae) leaf flushing occurred many times a year and was not predictable. Late dry season leaf flushing was observed in the deciduous trees of Sri Lanka, Southern Nigeria, Belgian Congo (Wright 1905, Coster 1923) while flushing in cooler months was recorded in Mexico and Osa peninsula of South East Costa Rica (Bullock and Solis-Magallanes 1990, Boinski and Fowler 1989). The actual factors responsible for this leaf flushing behavior are not yet obviously understood. Moisture stress has been considered as an important factor in leaf fall, since the phenomenon is generally observed during dry months (Njoku 1963, Hopkins 1970, Frankie et al. 1974). Wet season defoliation in trees is most likely due to unusual sensitivity of plants to low light intensities and high humidity that accompany beginning rainy season (Koriba 1958, Alvim 1964, Addicott and Lyon 1973).

Leaf fall may be total or partial depending on the taxa. In some actually deciduous taxa, all or most of the elderly leaves got abscised at a considerable time before the production of new ones so that the tree was bare for a period of weeks or a few months (*Terminalia* spp.). These taxa belong to the holo deciduous category recognized by Koriba (1958). In others such as *Sterculia urens* and *Ficus benghalen-*

sis and *Pongamia pinnata* leaf fall and leaf flushing processes overlap slightly in the similar tree. In certain others, only a portion of the tree was shed when some branches possessed intact leaves. This category called semi-deciduos by Koriba (1958) was also recorded in a few taxa. In mainly evergreens, old leaves abscised over a period of time throughout the year as and when they elderly, thus retaining a steady population of functional leaves all the time, but even in these cases there was often a period when leaf fall was substantial. All these categories of leaf fall patterns are very well predictable in the flora of several countries (Beard 1964, Boinski and Fowler 1989, Addicot 1978).

Flowering continued in different species throughout the year. However, peak period of flowering can be distinguished for most of the species in the month of March-April where plants like *Cassia alata*, *Saraca indica*, *Murraya koenigii*, *Alstonia scholaris*, *Cassia fistula*, *Azadirachta indica*, *Spondias pinnata* all these exhibited flower beginning in response to increasing distance end to end photoperiod. There is wide fluctuation in the phenology of a species from region to region due to changes in the climatic situation. Leaf fall : The average time of commencement and completion of leaf fall has been noted that this phenomenon starts from January and continues up to February. Maximum leaf fall occurred in January–February while the least was observed during September to December. Flowering : The observations about the time of beginning and completion of flowering of different species of medicinal plants are tabulated. The highest number of medicinal tree taxa was found to flower during the months of March and April and only a few were in bloom during the period October-December. The peak flowering period of tree taxa coincided with the hot and dry season March-April.

In present study seeds are shed down in cold season typically in December and develop in next spring season. In majority of plants the leaf falling period is cool season i.e. January and February, leaf flushing usually occurred in warm period i.e. in March and April, in rainy season i.e. in August flowering was at its peak and usually in September and October fruit maturation takes place. In small, in our learning area plants grow and goes up to flowering and fruiting

stage in humid and wet season and shed seeds which gain cold treatment in winter season and then once again grows up in favorable season.

Conclusion

The phenology study documentation on the circumstance of the flora of this vulnerable region we can know what actually occurrence to the biodiversity in the finest or accurate level. Floristic phenology works on micro level should not be continued to explore level or scientific population but we should make it open to the community about the position, circumstances of nature forest vegetation and awful effect of defeat of biodiversity and each and every one. Each step should be taken to keep the medicinal or other medicinal plant diversity of the region to region which is already in a critical level that can be necessary by observing the present percentage of forest, inclusion of species in (RET) Red Data Book or changed phenological behavior of the medicinal plant species. Understanding on floristic, phenology and reproduction systems for any forest ecosystem constituting medicinal plants taxa is very supportive in deciding strategies for responsible management of plant genetic resources including conservation activities. Periodicity of medicinal plants flowering and fruiting in tropical environments has received little attention until recently. The records so composed would be helpful for planning proper PGR management strategies in the Western Ghats region to maintain development without destruction.

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References

- Addicot TF (1978) Abscission strategies in the behavior of tropical trees. In : Tomlinson PB, Zimmermann MH (ed). Tropical trees as living systems.
- Addicott TF, Lyon JL (1973) Physiological ecology of abscission. In : Kozlowski TT (ed). Shedding of plant parts. New York : Academic Press.
- Ahmad K, Khan ZI, Ashraf M, Hussain MI, Aleem EH (2008)

- Status of plant diversity at Kufri (Soone Valley) Punjab, Pakistan and prevailing threats. *Pak J Bot* 40 (3) : 993—997.
- Ali SI (2008) Significance of Flora with special reference to Pakistan. *Pak J Bot* 40 (3) : 967—971.
- Alvim PT (1964) Tree growth periodicity in tropical climates. In : Zimmermann MH (ed). *Formation of Wood in Forest Trees*. New York : Academic Press.
- Annual Report (2001) Botanical Survey of India.
- Barman D, Nath N, Kishor Deka (2014) Phenology of some medicinal plant species of Goalpara District, Assam (India). *Sch Acad J Biosci* 2 (2) : 81—84.
- Bawa KS (1974) Breeding system of tree species in a tropical lowland community. *Evolution* 28 : 85—92.
- Beard JS (1964) The Mora forests of Trinidad, British West Indies. *J Ecol* 33 : 173—192.
- Bhatt RM, Purohit AN (1984) Morphophysiological behavior of 2 *Anaphalis* species from contrasting environments along an altitudinal gradient. *Ind J Pl Physiol* 27 : 130—137.
- Boinski S, Fowler NL (1989) Seasonal patterns in a tropical lowland forest. *Biotropica* 21 (3) : 223—233.
- Bullock SH (1985) Breeding systems in the flora of a tropical deciduous forest in Mexico. *Biotropica* 17 (4) : 287—301.
- Bullock SH, Solis-Magallanes JA (1990) Phenology of canopy trees of a tropical deciduous forest in Mexico. *Biotropica* 22 (1) : 22—33.
- Butola JS (2009) Propagation and Field trials using conventional methods, of some threatened medicinal plant species of Himachal Pradesh. PhD thesis. Forest Research Institute University, Dehradun, India.
- Butola JS, Badola HK (2007) Growth, phenology and productivity of *Dactylorhiza hatagirea* (D. Don) Soo, a critically endangered medicinal Orchid in Himalaya : Domestication compared with wild. *J Orchid Soc Ind* 20 : 37—43.
- Coster C (1923) Lauuberneuerung und andere periodische lebensprozesse in dem trockenen Monsungsgebiet Ost-Javas. *Annals du Jard Botanique Buitenzorg* 33 : 117—189.
- Croat TB (1979) The sexuality of Barro Colorado Island flora (Panama). *Phytologia* 42 : 319—348.
- Das Moumita, Pandey Archana SP (2007) Bajpai studies on phenological events of certain medicinal plants. *Int J Pl Sci* 2 (1) : 130—132.
- Devis PH, Heywood VH (1973) *Principles of angiosperm taxonomy*. Huntington, New York : Robert E. Kieger Publishing Co.
- Duraisamy Suresh, Palsumy Subramaniam (2010) Phenological observation and population dynamics of six uncommon medicinal plants in the grasslands of Nilgiris, Western Ghats, India. *Maejo Int J Sci Technol* 4 (02) : 185—192.
- Flores S, Schemske DW (1984) Dioecy and monoecy in the flora of Puerto Rico and the Virgin Island : Ecological Correlates. *Biotropica* 16 (2) : 132—139.
- Frankie GW, Baker HG, Opler PA (1974) Comparative inflammatory studies of trees in tropical West and dry forests in lowlands of Costa Rica. *J Ecol* 62 : 881—919.
- Freeman DC, Harper KT, Ostler K (1980) Ecology of plant dioecy in the intermountain region of Western North America and California. *Oecologia* 44 : 410—417.
- Gamble JS (1935) *The Flora of the Presidency of Madras*. Adlard & Son, Ltd, London.
- Hamann A (2004) Flowering and fruiting phenology of a philippine submontane rain forest climate factors as proximate and ultimate causes. *J Ecol* 92 : 24—31.
- Henry AN, Chitra V, Balakrishnan N (1987) *Flora of Tamil Nadu, India*. Botanical Survey of India, Coimbatore, pp 2.
- Hopkins B (1970) Vegetation of the Olokemeji forest reserve, Nigeria VI. The plants on the forest site with special reference to their seasonal growth. *J Ecol* 58 : 765—793.
- Jain SK (1963) Studies in Indian Ethnobotany plants used in by the tribes of Madhya Pradesh. *Bull Regional Res Lab* 1 : 126—129.
- Koriba K (1958) On the periodicity of tree-growth in the tropics, with reference to the mode of branching, the leaf-fall, and the information of the resting bud. *Gardens Bull Singapore* 17 : 11—81.
- Manske LL (2006) *Western snowberry biology*. Annual report North Dakota State University Dickinson, Research Extension Center.
- Matthew KM (1991) *An Excursion Flora of Central Tamil Nadu, India* Oxford and IBH Publishing Co, New Delhi, pp 47.
- Nair NC, Henry AN (1983) *The Flora of Tamil Nadu* Botanical Survey of India, pp 1.
- Nayar MP, Sastry ARK (1987, 1988, 1990) *Red data book of India plants*. Three volumes. Botanical Survey of India, Kolkata, India.
- Njoku E (1963) Seasonal periodicity in the growth and development of some forest trees in Nigeria I. Observations on mature trees. *J Ecol* 51 : 19—26.
- Oni PI (2001) Breeding systems in *Parkia biglobosa* (Jacq.) Benth ; an indigenous fruit resource utilization in Nigeria. *J Trop Resour* 17 : 1-9.
- Oni PI (2004) Initial evaluation of *Parkia biglobosa* (Jacq.) Benth provenances from West Africa countries. Regional Conference on Plant Genetic Resources and Food Security in West and Central Africa held at IITA (26th—30th, April 2004), pp 108—115.
- Oni PI (2010) Ethno-botanical survey of a fallow plot for medicinal plants diversity in Idena, Ijebu Ode. South Western Nigeria. *J Med Pl Res* 3 (10) : 45—52.
- Patel DK (2014) Herbaceous medicinal and aromatic plants diversity and introductions in herbal garden for *ex-situ* conservation. *Int J Herbal Med* 2 (3) : 17—20.
- Richards AJ (1986) *Plant Breeding Systems*. London : George Allen and Unwin.
- Sanz-Cortes F, Martinez-Calvo J, Badenes ML, Bleiholder H, Llacer G, Meier U (2002) Phenological growth critically endangered medicinal Orchid in Himalaya; Domestication compared with wild. *J Orchid Soc Ind* 20 (1-2) : 37—43.
- Sefidkon F, Kalvandi R, Atri M, Barazandeh MM (2003) Contribution for the Characterization of *Thymus Eriocalyx* Chemotypes. The international Magazine for Cosmetics and Fragrances.
- Sivaraj N (1991) Phenology and reproductive ecology of angiosperm taxa of Shervaroy hills (Eastern Ghats, South India). PhD thesis. Bharathidasan University, Tiruchirapalli, Tamil Nadu.
- Sivaraj N, Krishnamurthy KV (1989) Flowering phenology in the vegetation of Shervaroy, South India. *Vegetatio*

- 79 : 85—88.
- Sivaraj N, Kamala V, Pandravada SR, Sunil N, Elangovan M, Babu SB, Chakrabarty SK, Varaprasad KS, Krishnamurthy KV (2014) Floristic ecology and phenological observations on the Medicinal flora of Southern Eastern Ghats, India. *J Med and Arom Pl* 5 (2) : 5—22.
- Sundriyal RC, Joshi AP, Dhasimana R (1987) Phenology of high altitude plants at Tungnath in the Garhwal Himalaya. *Trop Ecol* 28 : 289—299.
- Van Schaik CP, Terborgh JW, Wright SJ (1993) The phenological of tropical forest : Adaptive significance and consequences for primary consumers. *Ann Rev Ecol Syst* 24 : 353-377.
- Vashistha RK, Nautiyal BP, Nautiyal MC (2006) Conservation status and morphological variations between populations of *Angelica glauca* Edgew. and *A. Archangelica* Linn. in Garhwal Himalaya. *Curr Sci* 91 : 1537—1542.
- Wright MH (1905) Foliar periodicity of endemic and indigenous trees in Ceylon. *An Royal Bot Gardens, Peradeniya* 2 : 415—517.
- Zhang X, Friedi MA, Schaaf CB, Strahler AH, Liu Z (2005) Monitoring the response of vegetation phenology to precipitation in Africa by coupling MODIS and TRMM instruments. *J Geophysic Res Atmospheres*,10: D12103, doi : 10.1029/2004JD005263.
- Zhang G, Song Q, Yang D (2006) Phenology of *Ficus racemosa* in Xishungbanna, South West China. *Bitropica* 38 : 334—341.