

Comprehensive Evaluation of Rice Genotypes for Salt Tolerance: *In vitro* Screening, Association Studies and Principal Component Analysis

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Received 26 June 2024, Accepted 25 September 2024, Published on 15 November 2024

ABSTRACT

Rice is an important staple crop of most of the people around the world. The production of rice is affected mainly by salinity in coastal lines. In the present study correlation and path analysis and principal component analysis is done for eight yield attributing traits to find out the genotypes for saline tolerance and decide on what trait basis the selection could be done. Correlation analysis showed that essential yield-related traits are plant height, panicle length, total number of tillers per plant, number of productive tillers per plant, number of grains per plant, and 1000 seed weight as they are positively significantly correlated with grain yield. Path analysis showed that all the traits had positive indirect effect on each other's except days to 50% flowering and plant height indicating

these two characters do not contribute much on yield. The highest positive direct effect on grain yield was obtained by number of grains per plant followed by 1000 seed weight and number of productive tillers per plant. Principal component analysis was performed for eight traits which showed PC1 and PC2 has eigen value greater than 1. The eigen value of PC1 and PC2 are 3.24 and 1.16 respectively. Percentage of variance for the two factors are 40.5% and 14.5% together accounting 55% of variability of the genotypes used for the study. The genotypes Kuzhi adichan, Kalanamak, Kalurundai samba, Vaikunda, Kalarpalai, Kottara samba, Illapai poo samba and Palkodai Valai which were tolerant to salinity and the genotypes ADT 37, ADT 39, ADT 42, ADT 43, ADT 45, ADT 47 which were susceptible to salinity were identified from PCA biplot and can be used for further breeding program.

Keywords Correlation analysis, Path analysis, Eigen value, Principal component analysis, PCA biplot.

INTRODUCTION

Rice (*Oryza sativa*) an edible starchy grain which is an important crop of the family Poaceae. It serves as a staple crop for two third of the world's population (Mohammadi-Nejad *et al.* 2010). The daily calorie requirement of upto 70% for most of the people around the world is provided by rice. Due to unforeseen conditions in global climatic condition rice production is severely affected by abiotic conditions such as salinity, drought and flood. One of the main issues af-

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fecting rice production globally, particularly in coastal areas, is salinity (Reddy *et al.* 2017). When salinity is present during pollination and fertilization, it affects rice sterility as well as the growth and development of the plant as well as its ability to adapt to stress (Pearson George and Leon 1959). Muhammad and Tomosaburo (1977) found that salinity causes panicle sterility in only some rice varieties, suggesting some genetic control (Khatun *et al.* 1995). Salinity delays heading in rice, which negatively affects a number of yield components (Grattan *et al.* 2002). Salinity stress has an impact on a number of plant functions, including flowering stage, spikelet count, percentage of sterile florets, leaf size, shoot growth, shoot and root length, shoot dry weight and shoot fresh weight (Muhammad and Nudrat 2009, Hakim *et al.* 2010, Gupta and Bingru 2014). Salinity can cause osmotic stress akin to physiological drought, and elevated salt accumulation in soils makes it harder for plants to absorb water and nutrients (Verslues Paul *et al.* 2006). Elevated salinity diminishes the flowering stage's pollen viability, which ultimately impacts grain output (Khatun and Flowers 1995, Singh *et al.* 2004). The degree of salt tolerance and the concentration of salt determine how differently different genotypes grow in response to salinity (Eynard *et al.* 2005). Developing crops that are resistant of high salinity and comprehending the mechanisms underlying this stress can help boost food supply. Salinity tolerant crops can be developed by the improvement of known varieties. The primary step in any crop improvement program is to evaluate and characterize the available genotypes for genetic variability. Correlation study helps to understand the association between yield and yield attributes, which ultimately helps in the realization of higher crop productivity. Path analysis helps to measure the direct and indirect contributions of independent variables to the dependent ones. It splits the correlation coefficient into two components, i.e., direct and indirect effects. Correlation and path analysis will describe the extent of association between seed yield and its attributes and also indicate the relative importance of their direct and indirect effects. The knowledge of existing genetic variability among the genotypes for different characters and the estimation of character association is important to make the selection program effective (Mohanty *et al.* 2024). The widely held belief that genetic variety

exists in the gene pool is a necessary condition for implementing a productive and worthwhile breeding strategy (Patel *et al.* 2022). Characterization of this existing variability and realignment of characters in them through selective breeding might go a long way in meeting the existing and emerging challenges that threaten the world food security (Vanaja and Babu 2006). Principal component analysis is one of the important tools used for identifying the plant characters that categorize the distinctiveness among the promising genotypes. Crop species vary on a regular basis, PCA aids in the removal of redundancy from data sets (Maji and Saibu 2012 and Ramakrishnan *et al.* 2014). In the present study correlation and path analysis and principal component analysis are done to evaluate the association of various characters to grain yield under salt stress in 50 genotypes of *O. sativa*.

MATERIALS AND METHODS

The present research work was carried out during the *kharif* 2022-2023 at Annamalai University, Department of Genetics and Plant Breeding, Annamalai Nagar, Tamil Nadu, India. The seed material for the present study was collected from various regions of Tamilnadu during 2021-2022. Totally 50 seed materials were collected from various agro climatic zones which includes indigenous rice varieties and commercial varieties. The collected germplasms were stored in the laboratory at 4°C. The list of genotypes used for the study were listed in the (Table 1).

The 50 rice genotypes were used for screening at the field condition and lab condition. Field was thoroughly prepared and levelled before sowing and before transplantation. During the season 50 rice genotypes were sown in the raised nursery beds. The experimental plot was laid out using Randomized Block Design (RBD) with 3 replications. Twenty-five days old seedlings were transplanted in the main field. All the standard agronomic practices recommended were followed to each plot and raised as a healthy crop and plant protection measures are done.

Evaluation of rice (*Oryza sativa* L.) genotypes for salt tolerance has become essential due to the increasing salinity stress in rice-growing areas. *In*

Table 1. List of genotypes used for the study.

Sl. No.	Genotype	Sl. No.	Genotype
1	Kuzhi adichan	26	ADT37
2	Tirupathi saram	27	ADT39
3	Marathundi	28	ADT40
4	Gobinth bhog	29	ADT41
5	Mutrilum sannam	30	ADT42
6	Samba mosanam	31	ADT43
7	Anai komban	32	ADT44
8	Vaikunda	33	ADT45
9	Ottatam	34	ADT46
10	Kalurundai samba	35	ADT47
11	Illapai poo samba	36	ADT48
12	Karuvachi	37	ADT49
13	Chinnar	38	ADT50
14	Kalajera	39	CO-50
15	Kalanmak	40	CO-51
16	Katti samba	41	BPT-5202
17	Pal kodai valai	42	BPT-5204
18	Anandanoor sanna	43	NLR - 3083
19	Milagu samba	44	NLR - 3449
20	Kottara samba	45	PS226
21	Manjal ponni	46	AC-39000
22	Kalarpalai	47	CR 1009
23	Poong karr	48	AC 35534
24	Kamban samba	49	TRY -1 (HTC)
25	ADT36	50	IR64 (HSC)

in vitro screening methods, association studies and principal component analysis are widely used techniques for assessing salt tolerance in rice genotypes. *In vitro* screening provides a quick and efficient way to evaluate the response of rice plants to salt stress by exposing them to controlled salt concentrations. Association studies involve studying the correlation between genotypic variations and salt tolerance traits in a large population of rice genotypes, which helps identify the genetic markers associated with salt tolerance. Furthermore, principal component analysis is a statistical tool that allows for the identification of the most influential traits contributing to salt tolerance. By applying these three approaches in combination,

reliable and accurate evaluation of rice genotypes for salt tolerance can be achieved, leading to the development of improved salt-tolerant varieties and ultimately ensuring food security in areas affected by salinity stress.

In vitro screening for salinity

All the 50 rice accessions were raised separately under normal and saline condition for seedling stage salinity tolerance using a hydroponic system under laboratory. The weather conditions that prevailed during the period were minimum temperature of 22°C to 24°C, maximum temperature of 30°C to 31°C, natural sunlight and relative humidity of 65% to 70%. The seeds were sterilized and placed in the germination paper and incubated at 27°C for 48 hrs to germinate. One well pre-germinated seeds were sown per hole on storiform floats with a net bottom and it was suspended on the plastic tray with Yoshida nutrient solution (Yoshida and Parao 1976). The radical should be inserted through the nylon mesh so as to prevent the damage to the pre-germinated seeds. Yoshida nutrient solution was prepared by using various chemical reagents containing macro and micro nutrients as required (Table 2).

Correlation and path analysis

Observations were recorded during the crop growth and at harvest time for the characters viz., Days to fifty per cent flowering, Plant height, Panicle length, Number of tillers per plant, Number of grains per panicle, Thousand seed weight and Grain yield and the data was subjected to statistical analysis. Statistical analyses for the above characters were done following Singh and Chaudhary (1977) for correlation coefficient and Dewey and Lu (1959) for path analysis.

Table 2. Mean value for saline analysis under laboratory condition.

Genotype	SL	RL	TSL	G%	FSW	DSW	SV -1	SES score
1	15.39	25.3	40.69**	89.51**	0.103**	0.104**	3642.40**	1
2	12.48	12.97	25.45	51.38	0.037	0.015	1311.13	3
3	8.55**	16.71	25.27	53.58	0.026	0.021	1354.43	5
4	10.27	6.44**	16.71	62.93	0.027	0.018	1052.18	7
5	11.6	13.71	25.31	63.63	0.023	0.023	1611.46	5
6	10.5	8.13*	18.63	70.16	0.028	0.023	1316.76	5
7	11.71	14.52	26.23	51.5	0.027	0.009	1352.17	7
8	19.96	20.35	40.32**	78.52	0.104**	0.090**	3171.6**	3

Table 2. Continued.

Genotype	SL	RL	TSL	G%	FSW	DSW	SV -1	SES score
9	12.35	18.62	30.97**	72.5	0.098**	0.053	2247.86*	5
10	15.51	21.43	36.94**	88.4**	0.107**	0.1	3263.37**	1
11	9.37*	9.45*	18.83	62.4	0.071**	0.060*	1196.39	7
12	21.76	6.8**	28.56*	50.33	0.080**	0.068*	1450.26	7
13	9.56	13.78	23.34	68.13	0.020	0.015	1592.81	5
14	10.62	12.51	23.14	62.3	0.028	0.023	1451.62	5
15	18.36	9.11*	27.47	91.41**	0.129**	0.103**	2511.86**	1
16	8.66**	13.5	22.16	44.42	0.098**	0.092	984.78	9
17	12.47	11.43	23.90	52.33	0.024	0.019	1259.11	7
18	10.00	13.93	23.94	54	0.019	0.019	1295.97	5
19	11.45	23.75	35.20**	34	0.019	0.009	1196.06	5
20	11.19	13.02	24.22	47.5	0.026	0.018	1153.02	7
21	15.95	7.03**	22.98	54	0.027	0.011	1242.78	7
22	10.47	18.03	28.5*	90.70**	0.105**	0.103**	2584.08**	1
23	14.44	25.33	39.77**	74.32	0.077**	0.067*	2958.21**	1
24	9.81	12.21	22.02	66.39	0.021	0.016	1449.74	5
25	10.26	11.18	21.45	77.3	0.024	0.018	1664.98	5
26	8.12*	9.51*	17.63	80.33	0.016	0.020	1413.99	7
27	11.10	9.95*	21.06	77.46	0.014	0.016	1631.99	7
28	9.90	6.60**	16.51	55.66	0.032	0.02	903.42	9
29	13.53	8.21**	21.74	74.33	0.019	0.017	1608.15	7
30	8.76*	10.28	19.04	65.39	0.021	0.028	1281.17	7
31	8.61*	12.15	22.76	74.68	0.020	0.006	1706.75	7
32	14.11	9.21	23.33	75.42	0.020	0.015	1748.88	7
33	12.48	13.12	25.60	35.45	0.021	0.028	907.26	9
34	13.83	8.64**	22.48	57	0.025	0.019	1275.77	7
35	16.73	8.18**	24.91	68.01	0.017	0.009	1686.65	7
36	13.89	12.01	25.91	71.87	0.02	0.017	1876.26	5
37	10.87	6.97**	17.85	55.66	0.024	0.015	1001.25	7
38	9.90	6.60**	16.51	76.33	0.032	0.02	1267.68	7
39	13.83	8.64**	22.48	57	0.025	0.019	1275.77	7
40	12.96	14.55	27.52	77.69	0.013	0.009	2120.61*	3
41	9.84	12.09	21.93	59.33	0.038	0.022	1315.91	9
42	10.00	12.10	22.11	74.33	0.019	0.017	1650.41	7
43	10.86	14.72	25.59	81.66*	0.03	0.017	2082.66	5
44	10.85	14.16	25.01	74.51	0.028	0.013	1864.66	7
45	9.76	13.30	23.07	74.41	0.026	0.018	1707.71	7
46	10.53	11.07	21.61	75.29	0.024	0.011	1652.07	9
47	10.75	11.80	22.56	82.25*	0.029	0.021	1853.34	5
48	10.23	10.82	21.06	70.33	0.026	0.016	1484.01	7
49	11.54	16.76	30.31**	90.26**	0.121**	0.158**	2544.36**	1
50	8.49*	13.43	21.93	33.6	0.031	0.091*	736.97	9
SE	0.86	0.87	1.37	5.33	0.005	0.011	157.69	
SEd	1.22	1.24	1.94	7.53	0.008	0.015	223.01	
CD (0.05)	2.42	2.46	3.84	14.92	0.016	0.031	441.56	
CD (0.01)	3.22	3.27	5.11	19.82	0.02	0.04	586.52	
	SL – Shoot length	RL – Root length	TSL – Total seedling length	G% - Germination percentage	FSW– Fresh seedling weight	DSW-Dry seedling weight	SV-1-Seedling vigour index 1	

Principal component analysis

The majority of the reported genotype changes were found to be attributed to certain plant features, which

were found using Principal component analysis. For eight quantitative traits, the mean values of 50 genotypes were employed. Principal components are generally estimated from correlation matrix or covariance

matrix. The data was analyzed using STAR software.

RESULTS AND DISCUSSION

Mean value for saline under laboratory condition is given in Table 2. All the genotypes showed varied response at seedling stage towards saline stress. The genotypes were scored from 1 (highly tolerant) to 9 (highly susceptible) (Rajabi Dehnavi *et al.* 2020). The resistant checks, TRY - 1 not show any symptoms under saline conditions and had normal growth and development (Rahman *et al.* 2016). While, the susceptible checks ADT - 40, ADT - 46, CO - 50 and BPT - 5202 showed whitish leaf tips, drying and rolling of leaves, and death of seedlings (Negrao *et al.* 2017). The genotypes like Kuzhi adichan, Kalamamak, Kalurundai samba kalarpalai and Kottara samba, (score of 1 and highly tolerant) also showed normal growth and development as resistant checks at seedling stage. The tolerant genotypes maintain ion homeostasis by ion uptake and compartmentalization of excess ions in the vacuole or older tissues (Hasegawa 2013). In addition to ion homeostasis, the tolerant genotypes accumulate large amount of osmolytes like proline, glycine betaine, polyols, and sugars to

maintain the osmotic balance (Saxena *et al.* 2013).

The results of the present study indicated genotypic correlation coefficient and phenotypic correlation coefficient were similar. Phenotypic and genotypic correlation analysis for eight characters of rice under salinity is given in Table 3. The grain yield was positively correlated with all the characters like panicle length, number of tillers per plant, number of productive tillers per plant, number of grains per plant and 1000 seed weight whereas days to 50% flowering and plant height are not significantly correlated with grain yield per plant at genotypic levels. This can be improved by selecting early maturing lines. The grain yield per plant had no significant correlation with days to 50% flowering (Islam *et al.* 2016). The associations between yield and plant height were revealed by Faysal *et al.* (2022). Phenotypic correlation was found significant between grain yield with all the characters except days to 50% flowering. Days to 50% flowering was correlated significantly only with panicle length at genotypic levels whereas at phenotypic levels it was correlated with plant height, panicle length, number of tillers per plant, number of grains per plant and 1000 seed weight. All the other characters were not

Table 3. Phenotypic and genotypic correlation analysis.

	Correlation coefficient	DFPF	PH	PL	NTPP	NPTPP	NGPP	1000 SW	GY
DFPF	r_g	1**	0.1791	0.373 **	0.180	-0.010	0.210	0.209	0.035
	r_p	1**	0.1791 *	0.3733 **	0.18 *	-0.0098	0.2106 **	0.2088 *	0.0353
PH (cm)	r_g	0.179	1**	0.5131 **	0.3618 **	0.1643	0.3078 *	0.4904 **	0.2262
	r_p	0.179*	1**	0.5122 **	0.3596 **	0.1633 *	0.3078 **	0.4887 **	0.2257 **
PL	r_g	0.373**	0.513**	1**	0.4802 **	0.2947 *	0.5044 **	0.3317 *	0.3535 *
	r_p	0.373**	0.5122	1**	0.478 **	0.2922 **	0.5032 **	0.3302 **	0.3523 **
NTPP	r_g	0.180	0.361**	0.480**	1**	0.3465 *	0.4133 **	0.4214 **	0.3924 **
	r_p	0.18*	0.356**	0.478**	1**	0.3441 **	0.4104 **	0.417 **	0.389 **
NPTPP	r_g	-0.010	0.164	0.294*	0.346*	1**	0.1818	0.209	0.3897 **
	r_p	-0.009	0.163*	0.292**	0.344**	1**	0.1813 *	0.2077 *	0.3863 **
NGPP	r_g	0.210	0.307*	0.504**	0.413**	0.818	1**	0.1636	0.4695 **
	r_p	0.210*	0.307**	0.503**	0.410**	0.181*	1**	0.163 *	0.4686 **
1000 SW	r_g	0.209	0.49**	0.331 *	0.421**	0.209	0.163	1**	0.3578 *
	r_p	0.208*	0.488**	0.503**	0.410**	0.181*	1**	0.163*	0.356 **
GY	r_g	0.035	0.226	0.353*	0.392*	0.389*	0.469**	0.357**	1**
	r_p	0.035	0.225**	0.352**	0.389**	0.386**	0.468**	0.356**	1**

DFPF- Days to fifty per cent flowering
 NTPP- Number of tillers per plant
 NGPP- Number of grains per plant
 PH- Plant height
 NPTPP- Number of productive tillers per plant
 1000 SW – 1000 Seed weight
 PL- Panicle length
 GY – Grain yield

Table 4. Path coefficient analysis of direct (diagonal) and indirect effects of various traits on grain yield.

		DFPF	PH	PL	NTPP	NPTPP	NGPP	1000 SW
DFPF	Genotypic	-0.11735	-0.01752	0.02506	0.01187	-0.00239	0.07922	0.05642
	Phenotypic	-0.11738	-0.01731	0.02544	0.01182	-0.00230	0.07910	0.05593
PH	Genotypic	-0.02102	-0.09778	0.03438	0.02374	0.03901	0.11578	0.13206
	Phenotypic	-0.02102	-0.09665	0.03491	0.02362	0.03834	0.11561	0.13090
PL	Genotypic	-0.04388	-0.05017	0.06702	0.03152	0.06996	0.18971	0.08934
	Phenotypic	-0.04382	-0.04951	0.06815	0.03139	0.06861	0.18900	0.08847
NTPP	Genotypic	-0.02122	-0.03537	0.03218	0.06564	0.08227	0.15545	0.11348
	Phenotypic	-0.02113	-0.03476	0.03258	0.06568	0.08079	0.15414	0.11170
NPTPP	Genotypic	0.00118	-0.01607	0.01975	0.02275	0.23741	0.06839	0.05628
	Phenotypic	0.00115	-0.01578	0.01991	0.02260	0.23479	0.06810	0.05563
NGPP	Genotypic	-0.02471	-0.03010	0.03380	0.02713	0.04316	0.37614	0.04405
	Phenotypic	-0.02472	-0.02975	0.03429	0.02695	0.04257	0.37559	0.04366
1000 SW	Genotypic	-0.02458	-0.04795	0.02223	0.02766	0.04961	0.06152	0.26931
	Phenotypic	-0.02451	-0.04723	0.02251	0.02739	0.04877	0.06122	0.26786

DFPF- Days to fifty per cent flowering

NTPP- Number of tillers per plant

NGPP- Number of grains per plant

PH- Plant height

NPTPP- Number of productive tillers

per plant

1000 SW – 1000 seed weight

PL- Panicle length

GY – Grain yield

significant with days to 50% flowering. The plant height was correlated genotypically with panicle length, number of tillers per plant, number of grains per plant and 1000 seed weight and phenotypically significant with all the traits. The report is also similar to the findings of Soujanya *et al.* (2020) and Srijan *et al.* (2016) which showed significant and positive associations between panicle length and plant height. The panicle length and number of tillers per plant was significantly correlated with all the traits at both phenotypic and genotypic levels. The number of productive tillers per plant is negatively correlated with days to 50% flowering at genotypic and phenotypic levels whereas it was not significant with 1000 seed weight at genotypic level. All other characters were significantly correlated with number of productive tillers per plant. The genotypic correlation was not significant between number of grains per plant and 1000 seed weight. The close relation of phenotypic and genotypic correlation coefficients is studied from the correlation study which indicates the masking effect of environment on character association. In the current study the yield attributing characters like number of tillers per plant, number of tillers per plant, number of grains per plant and 1000 seed weight had positive significant association at genotypic and phenotypic levels. These traits would be efficient for enhancing the yield, and the yield of rice in future

breeding programs would be increased. The correlation response in specific characters might lead to the selection of the predicted characters, and knowing the relationship between qualities would assist in the appropriate selection process (Goncalves *et al.* 2017). Generally, one or two desired traits could be selected at the same time, so a deep understanding of the effect of these traits on other characters was required. For example, yield improvement could be achieved by the genotype development with long panicle length and high number of panicles. Moreover, knowledge about the correlation between various characters with grain yield per plant, which is the most important target character, is required. By using this knowledge, this critical trait could be obtained by selecting easily observable traits. Our research results confirmed that essential yield-related traits are plant height, panicle length, the total number of tillers per plant, number of productive tillers per plant, number of grains per plant, and 1000 seed weight and they could be used as selection standards for rice grain yield enhancement.

Path coefficient analysis

Knowledge of the association among phenotypic characters and their effect on yield would be essential for selecting desired lines to be integrated with a breeding program and for releasing new varieties

Table 5. Eigen value, contribution of variability and factor loadings for the Principal Component axes.

	PC1	PC2	PC3	PC4	PC5	PC6	PC7	PC8
Eigen value	3.239217	1.156857	0.940538	0.750557	0.657584	0.559769	0.371391	0.324086
Variance percent	40.49022	14.46071	11.75673	9.381965	8.219805	6.99711	4.642386	4.051079
Cumulative variance percent	40.49022	54.95093	66.70765	76.08962	84.30942	91.306	95.94892	100
Standard deviation	1.7998	1.0756	0.9698	0.86635	0.8109	0.74818	0.60942	0.56929
Proportion of variance	0.4049	0.1446	0.1176	0.09382	0.0822	0.06997	0.04642	0.04051
Cumulative proportion	0.4049	0.5495	0.6671	0.7609	0.8431	0.91307	0.95949	1.000005
DFF	0.208214	0.645454	-0.30179	0.512899	-0.297	0.13085	0.130333	0.252137
PH	0.366888	0.256249	0.404559	-0.32343	0.446661	0.254773	0.177833	0.485196
PL	0.435126	0.201379	-0.19315	0.015522	0.369648	0.085149	-0.64744	-0.41169
NTPP	0.408822	-0.09693	0.04385	0.081421	-0.01901	-0.86566	-0.05038	0.250289
NPTPP	0.276329	-0.51633	0.036479	0.649585	0.329273	0.213209	0.278039	-0.05064
NGPP	0.36869	-0.05143	-0.55085	-0.42177	-0.00763	0.002016	0.553939	-0.27054
1000 SW	0.350641	0.115709	0.621422	0.015227	-0.44705	0.041319	0.155691	-0.50146
GY	0.361014	-0.43028	-0.12288	-0.1593	-0.51667	0.337901	-0.34824	0.376373

PH- Plant height

NPTPP-Number of productive tillers

per plant

1000 SW – 1000 Seed weight

PL- Panicle length

GY – Grain yield

NTPP- Number of tillers per plant

NGPP- Number of grains per plant

(Dhavaleshvar *et al.* 2019). The correlation between two factors could be divided into indirect and direct impact through other factors based on path coefficient analysis. The direct and indirect impacts among yield and yield-related components could be positive or negative, but the direct impact of that particular character and indirect impact via other components could be pointed out. Therefore, the determination of the path coefficient is crucial, so the correlation of these impacts could be observed partially and revealed the root and relationship that exist between yield-related elements and yield. In the present study, path coefficient analysis was carried out for eight traits to find their correlation with grain yield per plant under saline conditions (Table 4). Among the characters studied panicle length, number of tillers per plant, number of productive tillers per plant, number of grains per plant and 1000 seed weight had direct positive effect on grain yield per plant. The highest positive direct effect on grain yield was obtained by number of grains per plant followed by 1000 seed weight and number of productive tillers per plant. Positive direct effects of various traits on grain yield reported in the present research agree with the findings of Yadav *et al.* (2010) for the number of tillers per plant, Faysal *et al.* (2022) for test weight, Saleh *et al.* (2020) for 1000 grain weight. The negative effect was obtained

by days to 50% flowering and plant height. These two characters do not have any effect on improving grain yield. Our findings are against the findings of Muthuvijayaragavan and Murugan (2017) where they reported a negative direct effect of panicle length, number of tillers per plant and number of productive tillers per plant on grain yield per plant respectively.

The highest positive indirect effect on grain yield per plant via days to 50% flowering is number of grains per plant whereas the lowest indirect effect was by number of tillers per plant. The negative indirect effect on grain yield via days to 50% flowering is plant height and number of productive tillers per plant. The plant height on grain yield had positive indirect effect in the order number of 1000 seed weight, number of grains per plant, number of productive tillers per plant, panicle length and number of tillers per plant. It has indirect negative effect on days to 50% flowering. All the traits had positive indirect effect on each other's except days to 50% flowering and plant height indicating these two characters do not contribute much to yield. The highest positive indirect effect on grain yield via panicle length, number of tillers per plant, number of productive tillers per plant and 1000 seed weight, number of grains per plant. Number of grains per plant had highest positive indirect effect

via 1000 seed weight. Our results agreed with that of Reetisana *et al.* (2022) and Aarhi *et al.* (2019) which revealed that days to 50% flowering had a negative indirect impact on grain yield per plant via panicle length. Saleh *et al.* (2020) also reported that the indirect impact of plant height on grain yield via panicle length which was negative.

In this research, the indirect and direct effects of yield contributing factors and yield were calculated. The relationship between the effect and a causal factor was similar to its direct effect, so the true relationship was explained and plant breeder could be selected directly through these traits. However, the positive correlation could be caused by indirect effects in the case of the direct effect were negligible or negative. In the present study, in the selection process, the other factors were involved simultaneously. Besides, some restrictions could be used to limit the unexpected indirect effects in order to utilize the direct effect when the direct effect was strong and positive but the correlation was negative. For improvement of grain yield, based on the correlation and coefficient data, test weight, number of productive tillers per plant, number of grains per plant, number of productive tillers per plant and panicle length were essential factors, which should be considered as selection criteria. Overall, among the genotypes used, grain yield per plant showed a highly significant and positive correlation with its component traits like test weight, number of productive tillers per plant, number of grains per plant, number of productive tillers per plant and panicle length while it exhibited a strong negative correlation with plant height and days to 50% flowering. Therefore, a separated but simultaneous selection for enhancement of these traits could be executed. The results also revealed that the total number of tillers per plant had maximum positive direct effects as well as indirect effects on other traits, which suggested that selection based on this trait for grain yield would be most effective strategy. Additionally, the role of number of panicles had maximum positive effect on grain yield that could be used for selection (Balasubramanian and Vennila 2022).

Principal component analysis

The main purpose of principal component analysis is

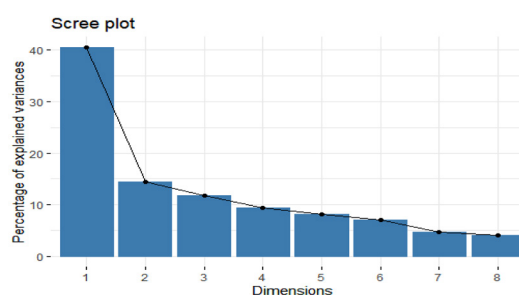


Fig.1. Screen plot showing eigen value and cumulative variability with their respective PCs.

reducing the dimension of a large dataset, increasing interpretability but at the same time minimizing information loss (Patel *et al.* 2022). Eigen vector values, percentage of variance and the cumulative percentage are presented in Table 5. In the present study PC1 and PC2 has eigen value greater than 1. The eigen value of PC1 and PC2 are 3.24 and 1.16 respectively. Percentage of variance for the two factors are 40.5% and 14.5% together accounting 55 % of variability of the genotypes used for the study. Similar type of finding also reported by Shaibu and Uguru (2017) and Salem *et al.* (2021). Rotated component matrix showed that PC1 exhibited highest variability 40.5% with highly loaded character viz panicle length, number of tillers per plant, number of grains per plant, 1000 seed weight and grain yield. It shows that PC1 is mainly dependent on panicle length and number tillers per plant. PC2 exhibited variance of 14.5 % with highly loaded for days to 50% flowering. Similar type of finding also reported by Sanni *et al.* (2012), Radhamani *et al.* (2015) and Lakshmi *et al.* (2019). The Principal component analysis revealed a wide range of scores for the eight quantitative traits, indicating a significant degree of variation. The analysis explained the variance structure through a few linear combination of the variables, proportion of variability. Thus, an accurate image of the component qualities that are providing the most variability is obtained based on factor loadings. Scree plots are graphs that display the eigen values in order of greatest to smallest as well as the percentage of variability in terms of primary component and eigen value. PC 1 showed 40.5% variability with eigen value 3.24 and then the graph gradually declines for another PC. Steep curve followed by bend and then straight found for

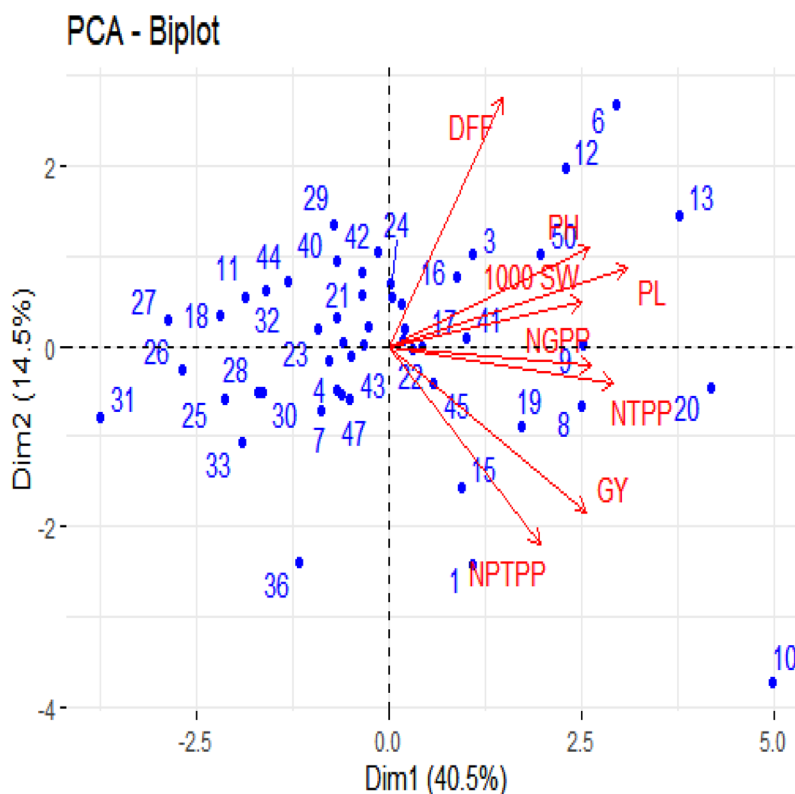


Fig. 2. Biplot analysis for 50 germplasm of rice with 2 PCs.

different PCs. The graph (Fig. 1) provided a concise explanation of the variability, which peaked for PC1. The distribution and kind of diversity for both the genotypes and the variables were described by the biplot diagram (Fig. 2) that shows the relationship between PCs 1 and 2. Nearly every genotype and variable showed a great degree of variance, as the loading plot illustrated. Ravi *et al.* (2018) observed similar reports.

The Biplot diagram (Fig. 2) revealed significant heterogeneity among genotypes and between parameters. In this regard, the study will be useful in finding the variability-contributing characteristics and selecting appropriate genotypes for breeding and use in crop development for yield-related traits. The results of the Biplot analysis are shown in Fig. 2. The results indicated that the two PCs were separated into two

clusters. The result of box plot revealed that cluster 1 was mainly characterized by plant height, number of tillers per plant and number of productive tillers per plant. The cluster 1 consisted of genotypes Kuzhi adichan, Kalanamak, Kalurundai samba, Vaikunda, Kalarpalai, Kottara samba, Illapai poo samba and Pal kodai valai. The cluster was characterized by 1000 seed weight and grain yield. The cluster 2 consists of genotypes ADT 37, ADT 39, ADT 42, ADT 43, ADT 45, ADT 47. Since the present study is done under salt stress condition we must select genotypes from 1st cluster for parents for saline breeding and genotypes from 2nd cluster as yield improvement. In the present study PCA revealed high level of genetic variation among the accession and the variable contributing the diversity. So, the result will be useful in choosing the parent in hybridization program for saline tolerance rice variety with higher yield performance.

REFERENCES

- Aarathi M, Vennila S, Palaniraja K (2019) Studies on genetic variability, correlation and path analysis in rice (*Oryza sativa* L.) under natural saline condition. *The Pharma Innovation Journal* 8 (7) : 528—532.
- Balasubramanian M, Vennila S (2022) Genetic diversity, correlation and path co-efficient for yield and yield associated traits in rice (*Oryza sativa* L.). *Crop Research* 57 (5 & 6) : 420—426.
https://doi.org/10.31830/2454-1761.2022.890
- Dewey DR, Lu KHA (1959) Correlation and path coefficient analysis of components of crested wheat grass seed production. *Agronomy Journal* 51 : 515—518.
https://doi.org/10.2134/agronj1959.00021962005100090002x.
- Dhavaleshvar M, Malleshappa C, Dushyanth-Kumar BM (2019) Variability, correlation and path analysis studies of yield and yield attributing traits in advanced breeding lines of rice (*Oryza sativa* L.). *International Journal Pure Applied Bioscience* 7 : 267—273.
DOI: <http://dx.doi.org/10.18782/2320-7051.7308>
- Eynard A, Lal R, Wiebe K (2005) “Crop response in salt-affected soils.” *Journal of Sustainable Agriculture* 27 (1) : 5—50.
https://doi.org/10.1300/J064v27n01_03
- Faysal ASM, Ali L, Azam MG, Sarker U, Ercisli S, Golokhvast KS (2022) Genetic variability, character association and path coefficient analysis in transplant aman rice genotypes. *Plants* 11: 2952.
https://doi.org/10.3390/plants11212952
- Goncalves DL, Barelli MAA, Oliveira TC, Santos PRJ, Silva CR, Poletine JP (2017) Genetic correlation and path analysis of common bean collected from Caceres Mato Grosso state, Brazil. *Ciencia Rural* 47: e20160815.
https://doi.org/10.1590/0103-8478cr20160815
- Grattan SR, Zeng L, Shannon MC, Roberts SR (2002) Rice is more sensitive to salinity than previously thought. *California Agriculture* 56 (6) : In press.
- Gupta Bhaskar, Bingru Huang (2014) “Mechanism of salinity tolerance in plants : Physiological, biochemical and molecular characterization.” *International Journal of Genomics*. 1 : 701596.
https://doi.org/10.1155/2014/701596
- Hakim MA, Juraimi AS, Begum M, Hanafi MM, Mohd R, Ismail Selamat A (2010) “Effect of salt stress on germination and early seedling growth of rice (*Oryza sativa* L.)” *African Journal of Biotechnology* 9 (13) : 1911—1918.
DOI:10.5897/AJB09.1526
- Hasegawa PM (2013) Sodium (Na) homeostasis and salt tolerance of plants. *Environmental and Experimental Botany* 92 : 19—31.
https://doi.org/10.1016/j.envexpbot.2013.03.001
- Islam MZ, Khalequzzaman M, Bashar MK, Ivy NA, Haque MM, Mian MAK (2016) Variability assessment of aromatic and fine rice germplasm in Bangladesh based on quantitative traits. *The Scientific World Journal* 2796720.
https://doi.org/10.1155/2016/2796720
- Khatun Saleha, Flowers TJ (1995) “Effects of salinity on seed set in rice.” *Plant, Cell and Environment* 18 (1) : 61—67.
https://doi.org/10.1111/j.1365-3040.1995.tb00544.x.
- Khatun S, Rizzo CA, Flowers TJ (1995) “Genotypic variation in the effect of salinity on fertility in rice.” *Plant and Soil* 173 : 239—250.
https://doi.org/10.1007/BF00011461
- Lakshmi VI, Sreedhar M, Vanisri S, Anantha MS, Rao LS, Gireesh C (2019) Multivariate analysis and selection criteria for identification of African rice (*Oryza glaberrima*) for genetic improvement of indica rice cultivars. *Plant Genetic Resources* 17 (6) : 499—505.
DOI: <https://doi.org/10.1017/S1479262119000327>
- Maji AT, Shaibu AA (2012) Application of principal component analysis for rice germplasm characterization and evaluation. *Journal of Plant Breeding and Crop Science* 4(6) : 87—93.
DOI: 10.5897/JPBCS11.093
- Mohammadi-Nejad MG, Singh RK, Arzani A, Rezaie AM, Sabouri H, Gregorio GB (2010) Evaluation of salinity tolerance in rice genotypes. *International Journal of Plant Production* 4 (3) : 199—208.
- Mohanty Aseem Amitabha, Kar Rajesh Kumar, Pradhan Banshidhar, Mishra Tapash Kumar, Das Subhashree, Dash Asit Prasad, Lenka Biswajit, Pradhan Hansaraj, Sahu Dibyabharati, Behera Chinmayee, Mishra Alankrita, Maity Gopalina (2024) Correlation and path analysis for quantitative traits in greengram. *Ecology, Environment and Conservation* 30 : 67—71.
<http://doi.org/10.53550/EEC.2024.v30i01.012>
- Muhammad Akbar, Tomosaburo Yabuno (1977) “Breeding for saline-resistant varieties of rice: IV. Inheritance of delayed-type panicle sterility induced by salinity.” *Japanese Journal of Breeding* 27 (3) : 237—240.
https://doi.org/10.1270/jsbbs1951.27.237
- Muhammad Ashraf, Aisha Akram Nudrat (2009) “Improving salinity tolerance of plants through conventional breeding and genetic engineering : An analytical comparison.” *Biotechnology Advances* 27 (6) : 744—752.
https://doi.org/10.1016/j.biotechadv.2009.05.026
- Muthuvijayaragavan R, Murugan E (2017) Inter relationship and path analysis in F₂ generation of rice (*Oryza sativa* L.) under submergence. *International Journal of Current Microbiology and Applied Sciences* 6 (9) : 2561—2571.
- Negrão S, Schmoekel S, Tester M (2017) Evaluating physiological responses of plants to salinity stress. *Annals of Botany* 119 : 1—11.
https://doi.org/10.1093/aob/mcw191
- Patel Ashish, Sao Abhinav, Nair Sunil, Mandavi Jageshwar, Tamrakar Nishesh (2022) “Principal component analysis for eight quantitative traits in 55 indigenous rice germplasm (*Oryza sativa* L.)” *Pharma Innovation Journal* 11 (9) : 1201—1206.
- Pearson George A, Leon Bernstein (1959) “Salinity effects at several growth stages of rice.” *Agronomy Journal* 654—754.
- Radhamani T, Sassikumar D, Packiaraj D, Saraswathi R, Ravi-V (2015) Principal component analysis of variation among the induced mutants of rice (*Oryza sativa* L.). *Research on Crops* 16 (4) : 698—703.
DOI : 10.5958/2348-7542.2015.00098.4
- Rahman MA, Thomson MJ, Shah-E-Alam M, de Ocampo M, Egdane J, Ismail AM (2016) Exploring novel genetic sources of salinity tolerance in rice through molecular and physiological characterization. *Annals of Botany* 117 : 1083—1097.

- <https://doi.org/10.1093/aob/mcw030>
- Rajabi Dehnavi AM, Zahedi A, Ludwiczak S, Cardenas Perez, Piernik A (2020) Effect of salinity on seed germination and seedling development of sorghum (*Sorghum bicolor* (L.) Moench) genotypes. *Agronomy* 10 : 859.
<https://doi.org/10.3390/agronomy10060859>
- Ramakrishnan P, Babu C, Iyanar K, Manivannan N (2014) Principal component analysis for evaluation of Guinea grass (*Panicum maximum* Jacq.) germplasm accessions. *The Bioscan* 11 (4) : 2859—2862.
- Ravi Yugandhar P, Kota Suneetha Usha Kiran B, Sridhar M (2018) Principal component analysis for agro morphological and quality characters in germplasm of rice (*Oryza sativa* L.). *International Journal of Advanced Biological Research* 8 (2) : 268—273.
- Reddy INBL, Kim BK, Yoon IS, Kim KH, Kwon TR (2017) Salt tolerance in rice: Focus on mechanisms and approaches. *Rice Science* 24 (3) : 123—144.
<https://doi.org/10.1016/j.rsci.2016.09.004>
- Reetisana N, Sastry EVD, Renuka T, Julia T, Artibashisha HP (2022) Correlation and path coefficient analysis studies in rice hybrids (*Oryza sativa* L.) genotypes of Manipur. *Biological Forum – An International Journal* 14 (2) : 972—980.
- Saleh MM, Salem KF, Elabd AB (2020) Definition of selection criterion using correlation and path coefficient analysis in rice (*Oryza sativa* L.) genotypes. *Bulletin of the National Research Center* 44 : 1—6.
<https://doi.org/10.1186/s42269-020-00403-y>
- Salem KF, Saleh MM, Aldahak L, Elabd AB (2021) Assessment phenotypic diversity of rice (*Oryza sativa* L.) genotypes by multivariate analysis. *Journal of Arid Land Agriculture* 7 : 52—59.
doi: 10.25081/jaa.2021.v7.6516
- Sanni KA, Fawole I, Ogunbayo SA, Tia DD, Somado EA, Futakuchi K (2012) Multivariate analysis of diversity of land-race rice germplasm. *Crop Science* 52 (2) : 494—504.
<https://doi.org/10.2135/cropsci2010.12.0739>
- Saxena SC, Kaur H, Verma P, Petla BP, Andugula VR, Majee M (2013) Osmoprotectants: Potential for crop improvement under adverse conditions. In: *Plant Acclimation to environmental stress. Plant Acclimation to Environmental Stress*, pp 197—232.
https://doi.org/10.1007/978-1-4614-5001-6_9
- Shaibu AA, Uguru MI (2017) Application of multivariate analysis to access selected rice germplasm phenotypic diversity. *International Journal of Experimental Agriculture* 25 : 1—8.
<https://doi.org/10.9734/JEAI/2017/31662>
- Singh RK, Chaudhary BD (1977) *Biometrical Methods in Quantitative Genetic Analysis*. Kalyani Publ, New Delhi, India.
- Singh RK, Mishra B, Singh KN (2004) “Salt tolerant rice varieties and their role in reclamation program in Uttar Pradesh.” *Indian Farming* 2 : 6—10.
- Soujanya T, Hemalatha V, Revathi P, Prasad MS, Yamini KN (2020) Correlation and path analysis in rice (*Oryza Sativa* L.) for grain yield and its components. *The Journal of research PJTSAU* 48 : 88—92.
- Srijan A, Kumar SS, Raju CD, Jagadeeshwar R (2016) Character association and path coefficient analysis for grain yield of parents and hybrids in rice (*Oryza sativa* L.). *Journal of applied and Natural Science* 8 : 167—172
DOI: <https://doi.org/10.31018/jans.v8i1.768>
- Vanaja T, Babu LC (2006) Variability in grain quality attributes of high yielding rice varieties (*Oryza sativa* L.) of diverse origin. *Journal of Tropical Agriculture* 44 (1-2) : 61—63.
- Verslues Paul E, Agarwal Manu, Agarwal Surekha Katiyar, Zhu Jianhua, Zhu Jian-Kang (2006) “Methods and concepts in quantifying resistance to drought, salt and freezing, abiotic stresses that affect plant water status.” *The Plant Journal* 45 (4) : 523—539.
<https://doi.org/10.1111/j.1365-3113X.2005.02593.x>
- Yadav SK, Suresh BG, Pandey P, Kumar B (2010) Assessment of genetic variability, correlation and path Association in rice (*Oryza sativa* L.). *Journal of Biological Science* 18 (1) : 1—8.
- Yoshida S, Parao FT (1976) Climatic influence on yield and yield components of lowland rice in the tropics. *Climate and rice* 20 : 471—494.