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Effect of Polyethylene Glycol (PEG) on Seed Germination of Important *kharif* Pulse Crops of Kashmir Valley

Asma Majid, Gul Zaffar, F. A. Lone, Zahoor A. Dar, Showkat A. Waza, Parvaiz A. Sofi, Ajaz A. Lone

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ABSTRACT

Pulses form an important part of human diet as they are a potent source of proteins. Drought is one of the most important abiotic factors which adversely affect pulse production. Seed germination is considered as the first and foremost feature which directly affect the growth and yield of crop plants. To study the effect of PEG (polyethylene glycol) induced drought on germination of *kharif* pulse crops of Kashmir including cowpea, common bean and mung bean, an experiment was laid out under laboratory conditions

¹Senior Research Fellow, ²³Professor, ^{4,5}Associate Professor Division of Genetics and Plant Breeding, SKUAST Kashmir, Shalimar Srinagar 190025 J and K, India

F.A. Lone

Professor, Division of Environmental Sciences, SKUAST Kashmir, Shalimar Srinagar, J and K, India

Showkat A. Waza⁵

Assistant Professor, Mountain Crop Research Station, Sagam, (Anantnag), SKUAST Kashmir, J and K, India

Email: asmamajid173@gmail.com *Corresponding author

at SKUAST-K Shalimar. Different parameters including, length of radical, number of laterals, root biomass and total number of germinating seeds were measured under different concentrations of PEG-6000 viz., 0% (Control), 5%, 10% and 20%. All the parameters estimated across the genotypes showed progressive decline with the increase in PEG concentration from 0 to 20%. In common bean, French Yellow exhibited lesser reduction in all the germination parameters with an increase in concentration of PEG-6000. For cowpea, genotypes UDC-46 and C-37 showed lesser reduction in the traits recorded. Similarly in case of mung bean, KM-1046 and KM-1854 exhibited lesser reduction in different parameters with the increase in concentration of PEG-6000. These genotypes after further evaluation can be used in breeding programs for the development of drought tolerant varieties.

Keywords Drought, Abiotic factor, PEG, *Kharif* pulses, Cowpea.

INTRODUCTION

The major food requirement of human beings is being fulfilled by cereals and legumes. Legumes rank second after cereals in terms of food production and contribute about 33% of the protein needs. They have been recognized as a major source of vegetable protein rich in minerals and vitamins. Legumes are grown in all types of climatic conditions with wide range of soil types (Graham and Vance 2003). They

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Asma Majid^{1*}, Gul Zaffar², Zahoor A. Dar³, Parvaiz A. Sofi⁴, Ajaz A. Lone⁵

play a vital role in maintaining soil fertility by fixing atmospheric nitrogen. According to the projections of Intergovernmental Panel on Climate Change (IPCC), reduction in yield as a result of climate change related abiotic stresses will be more pronounced for pulse crops. Among the abiotic stresses, drought is one of the most important environmental factors that reduce growth, development and yield of crop plants (Verslues et al. 2006, Xiong et al. 2006, Trenberth et al. 2014, Zhao and Dai 2015). Therefore, drought serves as the most important constraint for pulse productivity and the current challenge is to reduce the yield gaps between drought and non-drought conditions (Waza et al. 2013a, Dharbale et al. 2019, Majid et al. 2020a). Transfer of drought tolerance from tolerant to susceptible genotypes serves as the most coherent approach to develop the drought tolerant varieties. Drought stress tolerance being a complex phenomenon involves clusters of gene networks. As such, despite of various efforts during the past few decades, identifying plant types for use in breeding programs to transfer drought resistance traits into high yielding cultivars has become a challenge for plant breeders (Serraj and Buhariwalla 2004, Waza et al. 2013b, Dar et al. 2021). To achieve this objective, the screening for sources of drought resilient is indispensable.

The first and foremost effect of drought stress is on germination of seeds (Harris et al. 2002, Duman 2006, Waza et al. 2014). For research purpose, the creation and maintenance of pure water potential in the environment of soil is almost a difficult job. In this regard, establishing conditions of dryness stress using different osmotic materials to create the osmotic potential is considered as one of the best methods to study the effects of dryness stress on germination. Among these substances, due to the simulation of natural environmental conditions, polyethylene glycol (PEG) has many applications and is widely used in vitro (Ibrahim et al. 2001, Umesh et al. 2015). Because this compound has a high molecular weight, it cannot pass through the cell wall and therefore it is used to regulate water potential in germination tests. The principal aim of this study is to investigate the effects of osmotic stress generated by PEG on germination characteristics of some elite pulse crops grown under Kashmir conditions. This is to screen the germplasm for drought stress at an early stage of plant growth.

Experimental details

This experiment was carried out during 2020 under laboratory conditions at Sher-e-Kashmir University of Agricultural Sciences and Technology of Kashmir (SKUAST-K). Twenty two pulse germplasm lines consisting of eleven common bean (*Phaseolus vulgaris*) genotypes viz., WB-9716, WB-1446, WB-934, WB-112, WB-22, DARS-10, R-15, French Yellow, Shopian local, Budgam local, Ganderbal local; six cowpea (*Vigna unguiculata*) genotypes viz., SKUASTC-407, UDC-45, UDC-46, C-37, C-46, C-5 and five mung bean (*Vigna radiata*) genotypes viz., SKUAST M-1047, Mung Local Budgam, KM-2241, KM-1046 and KM-1854 were evaluated in the present study. The design followed was Completely Randomized Design (CRD) with three replications.

Induction of water stress using PEG-6000

Water stress was induced using different osmotic potential levels [0 (control), 5, 10 and 20%] to evaluate the effect of PEG-6000 treatments on seed germination. For each level of stress, four seeds of each cultivar were selected and sterilized in sodium hypochlorite (1%) and then washed twice in distilled water. The seeds of pulse crops were germinated in petri dishes on 2 layers of filter paper in a germinator maintained at 25 °C and 75% humidity in darkness. These petri plates were visualized on daily basis, filter papers replaced if needed and PEG solution was added as per the requirement.

Recording of data

After seven days, the length of radicle (cm), number of laterals, radicle weight (g) and total number of seeds germinated was estimated. Length of radicle was estimated with the help of scale and graph paper. Number of laterals was visually counted with the help of magnifying glasses. Radicle weight was estimated with the help of digital weighing machine. Percentage of germinating seeds was determined by counting the number of seeds germinated out of total number of seeds.

Genotypes	Radical length			Number of laterals				Root biomass				
	0%	5%	10%	20%	0%	5%	10%	20%	0%	5%	10%	20%
WB-9716	4	3	2.5	1	7	5	4	3	0.08	0.06	0.04	0.01
WB-1446	22	12	7	5	58	21	5	4	0.94	0.54	0.14	0.06
WB-934	20.5	7	7	7	16	7	6	4	0.54	0.07	0.06	0.06
WB-112	15	15	10	1	26	25	3	1	0.62	0.41	0.31	0.02
WB-22	7	4	3	1	4	4	2	1	0.09	0.07	0.05	0.02
DARS-10	13.5	10.5	9	2	19	19	7	1	0.58	0.12	0.11	0.04
R-15	10	6.5	5	1	13	4	2	1	0.14	0.08	0.5	0
French Yellow	14	12	7.5	4	18	16	14	13	0.43	0.28	0.19	0.17
Shopian local	11	6	4	1	9	5	4	2	0.36	0.06	0.05	0.02
Budgam local	11	5	4.5	4	31	6	5	5	0.18	0.06	0.05	0.04
Ganderbal local	7	5	4	1	8	6	3	3	0.09	0.08	0.05	0.01
Mean	12.27	7.82	5.77	2.18	19.00	10.73	5.00	3.00	0.37	0.17	0.14	0.04
CD (p ≤0.05)	Genotype=0.625				Genotype $= 0.87$				Genotype=0.012			
	PEG levels=0.267				PEG levels $=0.32$			PEG levels=0.004				
	Genotype x PEG levels=1.23			1.23	Genotype x PEG levels=1.78			Genotype x PEG levels=0.02				

Table 1. Effect of osmotic potential induced by PEG - 6000 on length of radicle (cm), number of laterals and root biomass (g) of common bean genotypes.

RESULTS AND DISCUSSION

Genotypic variation for radical length, number of laterals and root biomass under different stress levels

Comparison of the data revealed that length of radical, number of laterals and root biomass shows different performance under different stress levels (Tables 1 - 3).

Under controlled conditions the length of radical was found to be highest in WB–1446 (22cm) in common bean, C-407 (15cm) in cowpea and KM-1046 and KM-1854 (20cm) in case of mung bean. Under

5% PEG, the length of radical was found to be highest in WB-112 (15cm) in common bean, C-37 (12cm) in cowpea and KM-1046 (16cm) in case of mung bean. For 10% PEG, the length of radical was found to be highest in WB-112 (10cm) in common bean, C-37 (7.5cm) in cowpea and KM-1046 and KM-1854 (12cm) in case of mung bean. Using 20% PEG, the length of radical was found to be highest in WB-934 (7cm) in case of common bean, C-37 (6.5cm) in cowpea and KM-2241 (10cm) in case of mung bean.

Under controlled conditions number of laterals was observed to be highest in WB-1446 (58) in common bean, SKUAST C-407(16) in cowpea and KM-1046 (28) in case of mung bean. For 5% PEG,

Table 2. Effect of osmotic potential induced by PEG - 6000 on length of radicle (cm), number of laterals and root biomass (g) of cowpea genotypes.

Genotypes	Radical length			Number of laterals				Root biomass				
	0%	5%	10%	20%	0%	5%	10%	20%	0%	5%	10%	20%
SKUAST C-407	15	8	4.5	3.5	16	6	3	1	0.31	0.28	0.24	0.21
UDC-45	10	8	7	1	4	3	2	1	0.24	0.22	0.11	0.01
UDC-46	9	8	7	5	4	3	3	2	0.24	0.12	0.09	0.06
C-37	14	12	7.5	6.5	9	7	6	4	0.36	0.26	0.13	0.09
C-46	13	7.5	4	2	6	4	2	2	0.35	0.05	0.03	0.02
C-5	8	7	5	1	5	3	1	1	0.16	0.09	0.04	0.01
Mean	11.50	8.42	5.83	3.17	7.33	4.33	2.83	1.67	0.28	0.17	0.11	0.07
CD ($p \le 0.05$) Genotypes=0.312 PEG levels=0.179 Construct a PEC levels=1.46		-1.46	Genotypes=0.210 PEG levels=0.10			Genotype=0.018 PEG levels=0.06						

Genotypes		Radical length				Number of laterals				Root biomass			
	0%	5%	10%	20%	0%	5%	10%	20%	0%	5%	10%	20%	
SKUAST M-1047	16	10	9	7	10	9	8	3	0.36	0.14	0.1	0.09	
Mung Local Budgam	15	12	6	0	16	9	6	0	0.38	0.33	0.06	0	
KM-2241	18	12	10	10	15	13	12	10	0.28	0.14	0.12	0.09	
KM-1046	20	16	12	8	28	25	16	14	0.39	0.25	0.16	0.12	
KM-1854	20	15	12	9	16	14	10	10	0.36	0.23	0.19	0.14	
Mean	17.8	13.0	9.80	6.80	17.0	14.0	10.4	7.40	0.35	0.22	0.13	0.09	
CD (p≤0.05)	Genotype=0.14 PEG levels=0.54 Genotype x PEG levels=1.24			Genotype=0.35 PEG levels=0.8 Genotype x PEG levels=1.41			Genotype=0.016 PEG levels=0.03 Genotype x PEG levels=0.06						

Table 3. Effect of osmotic potential induced by PEG - 6000 on length of radicle (cm), number of laterals and root biomass (g) of mung bean genotypes.

the number of laterals was found to be highest in WB-112 (25) in common bean, C-37 (7) in cowpea and KM-1046 (25) in case of mung bean. Using 10% PEG, the number of laterals was found to be highest in French Yellow (14) in common bean, C-37 (6) in cowpea and KM-1046 (16) in mung bean. Using 20% PEG, the number of laterals was found to be highest in French Yellow (13) in case of common bean, C-37 (4) in cowpea and KM-1046 (14) in case of mung bean.

Under controlled conditions, the root biomass was found to be highest in WB-1446 (0.94g) in common bean, C-37 (0.36g) in cowpea and KM-1046 (0.39g) in case of mung bean. For 5% PEG, the root biomass was found to be highest in WB-1446 (0.54g) in common bean, SKUAST C-407 (0.28g) in cowpea and KM-1046 (0.25g) in mung bean. Using 10% PEG, the root biomass was found to be highest in WB-112 (0.31g) in common bean, SKUAST C-407 (0.24g) in cowpea and KM-1854 (0.19g) in case of mung bean. On application of 20% PEG, the root biomass was found to be highest in French Yellow (0.17g) in common bean, SKUAST C-407(0.21g) in cowpea and KM-1854 (0.14g) in case of mung bean.

The effect of PEG in physiological processes has been reported in cowpea (Badiane *et al.* 2004, Jain and Saxena 2016), mung bean (Afzal *et al.* 2005, Jain *et al.* 2015), moth bean (Soni *et al.* 2011), black gram (Priyanka *et al.* 2013, Yadav *et al.* 2013) and common bean (Nunes *et al.* 2008, Silva *et al.* 2016, Majid *et al.* 2016). The *in vitro* culture techniques minimize confounding effects on account of lack of control in environmental variables due to defined nutrient media, controlled conditions and homogeneity of stress application. Polyethylene glycols (PEG) of high molecular weights have been used to simulate water stress in plants as non-penetrating osmotic agents, lowering the water potential in a way similar to soil drying (Larher *et al.* 1993, Majid *et al.* 2017).

Germination percentage of genotypes under different concentration of PEG-6000

In common bean, germination percentage of genotypes under different concentration of PEG-6000 is shown in Table 4. Under controlled conditions germination percentage was found to be highest in

 Table 4. Germination percentage of genotypes under different concentration of PEG-6000.

Genotypes	Germination percentage (%)							
	0% PEG	5% PEG Commo	10% PEG on bean	20% PEG				
WB-9716	75	50	25	25				
WB-1446	75	75	75	50				
WB-934	75	75	50	50				
WB-112	75	75	75	0				
WB-22	75	75	25	0				
DARS-10	75	75	75	50				
R-15	75	75	75	0				
French Yellow	100	100	75	50				
Shopian local	75	75	75	25				
Budgam local	75	75	75	50				
Ganderbal local	75	50	25	25				

Table 4. Continued.

Genotypes	Germination percentage (%)						
()% PEG	5% PEG	10% PEG	20% PEG			
	Common bean						
		Cowpea					
SKUAST C-407	100	100	75	50			
UDC-45	75	75	75	50			
UDC-46	100	100	100	75			
C-37	100	100	100	100			
C-46	75	75	50	25			
C-5	75	50	50	25			
		Mung bean					
SKUAST M-104	7 100	100	100	100			
Mung Local	100	100	100	0			
Budgam							
KM-2241	100	75	75	75			
KM-1046	100	100	75	75			
KM-1854	100	100	100	100			

French Yellow (100%) in case of common bean. While as SKUAST C-407, UDC-46 and C-37 (100%) showed highest germination percentage in case of cowpea. Further genotypes viz., SKUAST M-1047, Mung Local Budgam, KM-2241, KM-1046 and KM-1854 (100%) recorded highest and equal germination percentage in case of mung bean. Under 5% PEG, germination percentage was found to be highest in French Yellow (100%) in common bean, SKUAST C-407, UDC-46 and C-37 (100 %) in cowpea and SKUAST M-1047, Mung Local Budgam, KM-1046 and KM-1854 (100%) in case of mung bean. For 10% PEG, germination percentage was found to be highest in WB-1446, WB-112, DARS-10, R-15, French Yellow, Shopian local and Budgam local (75%) in common bean; UDC-46 and C-37 (100 %) in cowpea and SKUAST M-1047, Mung Local Budgam and KM-1854 (100%) in case of mung bean. On application of 20% PEG, germination percentage was found to be highest in WB-1446, WB-934, DARS-10, French Yellow and Budgam local (50%) in common bean, C-37 (100%) for cowpea and SKUAST M-1047 and KM-1854 (100%) in case of mung bean. According



Fig. 1. Promising genotypes of cowpea, common bean and mung bean under different PEG concentrations.

to Ayaz *et al.* (2000), decrease in seed germination under stress conditions is due to the occurrence of some metabolic disorders. Water deficit shows negative effect on germination of seed and the growth of seedlings. The adverse effect of water shortage on germination and seedling growth has been well reported in different crops (Mostafavi *et al.* 2011).

Genotypes exhibiting least reduction with increase in concentration of PEG–6000

In common bean, French Yellow exhibited lesser reduction in different growth parameters with the increase in concentration of PEG-6000 from 0% to 20%. Similarly in case of cowpea, genotypes UDC-46 and C-37 showed lesser reduction in the different growth traits. For mung bean, KM-1046 and KM-1854 exhibited lesser reduction in all the growth parameters with the increase in concentration of PEG-6000 (Fig. 1). The genotypes need to be further evaluated under green house and field conditions. According to the previous studies, induced water deficit by polyethylene glycol shows similar values to that observed under the field conditions (Thill *et al.* 1979, Majid *et al.* 2020b).

CONCLUSION

Keeping in view the above stated research findings, it can be concluded that some of the genotypes exhibited lesser reduction in different growth parameters with the increase in concentration of PEG-6000 from 0% to 20%. The genotypes include French Yellow in common bean, UDC-46 and C-37 in cowpea and KM-1046 and KM-1854 in mung bean. These genotypes after further evaluation can be used in breeding programs for the development of drought tolerant varieties.

REFERENCES

- Afzal A, Khan A, Khalil S, Abdullah (2005) Effects of polyethylene glycol concentrations and duration on the yield of mung bean. *Sarhad J Agric* 21: 171 -175.
- Ayaz FA, Kadioglu A, Urgut RT (2000) Water stress effects on the content of low molecular weight carbohydrates and phenolic

acids in Cienanthe setosa. Canadian J Pl Sci 80(2): 373-378.

- Badiane FA, Diouf D, Sané D, Diouf O, Goudiaby V, Diallo N (2004) Screening cowpea (Vigna unguiculata L. Walp) varieties by inducing water deficit and RAPD analyses. African J Biotechnol 3(3): 174-178.
- Dar MH, Bano DA, Waza SA, Zaidi NW, Majid A, Shikari AB, Ahangar A, Hossain M, Kumar A, Singh US (2021) Abiotic stress tolerance-progress and pathways of sustainable rice production. *Sustainability* 13(4): 2078.
- Dharbale BB, Prashant B, Mahesh V, Dhanashri D, Nigade D (2019) Climate change and drought: A review impact of climate change and drought on pulses crops in India. *Ind J* Agric Allied Sci 5(2): In press.
- Duman I (2006) Effects of seed priming with PEG or K3PO4 on germination and seedling growth in lettuce. *Pak J Biol Sci* 11: 923-928.
- Graham PH, Vance CP (2003) Legumes: Importance constraints to greater use. *Pl Physiol* 131: 872-877.
- Harris D, Tripathi RS, Joshi A (2002) On-farm seed priming to improve crop establishment yield in dry direct-seeded rice. In: Pandey S, Mortimer M, Wade L, Tuong TP, Lopes K, Hardy B (eds.). Direct seeding: Research strategies and opportunities, internati res institute, Manila, Philippines, pp 231-240.
- Ibrahim M, Zeid N, El–Semary A (2001) Response of two differentially drought tolerant cultivars of maize to drought stress. *Pak J Biol Sci* 4: 779–784.
- IPCC. Intergovernmental Panel on Climate Change. https://www.ipcc.ch.
- Jain K, Mishra MN, Sharma R, Bano DA, Waza SA (2015) Mutation induced variability in bread wheat (*Triticum aestivum* L. CVS. PBW-373, Raj-3765). *Environ Ecol* 33(1A): 297-302.
- Jain C, Saxena R (2016) Varietal differences against PEG induced drought stress in cowpea. Octa J Environm Res 4(1): 058-062.
- Larher Z, Silva H, Klessig DF (1993) Active oxygen species in the induction of plant systemic acquired resistance by salicylic acid. Science–AAAS–Weekly Paper Edition–including Guide to Scientific Information 262(5141): 1883-1885.
- Majid A, Dar SA, Wani SH (2016) Stability analysis for yield its attributes in fieldpea (*Pisum sativum L.*) under Kashmir conditions. J Pl Sci Res 32(2): 101-105.
- Majid A, Zaffar G, Lone FA, Dar ZA, Sofi PA, Lone AA (2020a) Effect of polyethylene glycol (PEG) induced drought stress on seed germination of lentil (*Lens culinaris*) genotypes of Kashmir valley. *SKUAST J Res* 22(2): 36-40.
- Majid A, Parray GA, Sofi NR, Shikari AB, Waza SA (2020b) Combining ability effects for various agro- morphological traits in rice under temperate conditions. *Curr J Appl Sci Technol* 39(38): 47-58.
- Majid A, Parray GA, Wani SH, Kordostami M, Sofi NR, Waza SA, Shikari AB, Gulzar S (2017) Genome editing and its necessity in agriculture. *Int J Curr Microbiol Appl Sci* 6(11): 5435-5443.
- Mostafavi KH, Sadeghi GH, Dadresan M, Zarabi M (2011) Effects of drought stress on germination indices of corn hybrids (*Zea mays* L.). *Int J Agric Sci* 1(2): 10–18.
- Nunes CMJ, Araújo SS, Marques SJ, Fevereiro P, Bernandes SAR (2008) Physiological responses of the legume model *Med*-

icago truncatula cv. *Jemalong* to water deficit. *Environ Exp Bot* 63: 289-296.

- Priyanka K, Jaiswal HK, Waza SA (2013) Identification of restorers an maintainers for wild abortive cytoplasmic male sterile lines in rice. *J Appl Biosci* 39(2): 121-122.
- Serraj R, Buhariwalla HK (2004) Crop improvement of drought resistance in pulses: Aholostic approach. *Ind J Pulses Res* 17(1): 1-13.
- Silva M, Willadino L, Deobrah Y, Santos C, Olivaires A (2016) Response of ricinis communis to *in vitro* water stress induced by polyethylene glycol. *Pl Growth Regul* 78: 195–204.
- Soni P, Rizwan M, Bhatt K, Mohapatra T, Singh G (2011) In vitro response of Vigna aconitifolia to drought stress induced by PEG 6000. J Agron Crop Sci 7: 108–121.
- Thill DC, Schimman RD, Appeby AP (1979) Osmotic stability of mannitol and polethylene glycol 20.000 solutions used as seed germination media. *Agron J* 71: 105–108.
- Trenberth KE, Dai A, Van SG, Jones PD, Barichivich J, Briffa KR, Sheffield J (2014) Global warming and changes in drought. *Nat Clim Change* 4: 17–22.
- Umesh, Jaiswal HK, Sravan T, Waza SA, Bhardwaj R (2015) Estimation of genetic variability, heritability and genetic advance for yield and quality traits in some indigenous Basmati rice (*Oryza sativa* L.) genotypes. *Int J Farm Sci* 5(4): 32-40.

- Verslues PE, Agarwal M, Katiyar AS, Zhu J, Zhu JK (2006) Methods concepts in quantifying resistance to drought, salt, freezing, abiotic stresses that affect plant water status. *Pl J* 45: 523–539.
- Waza SA, Rastogi NK, Sharma D, Verulkar SB, Bano DA (2013b) Characterization of Indira Sona variety of hybrid rice. *Bio*infolet- A Quart J Life Sci 10(3B): 1065-1066.
- Waza SA, Rastogi NK, Verulkar SB, Sharma D (2013a) Studies on genetic relatedness among various genotypes of rice (*Oryza sativa* L.) using SSR markers. *Oryza* 50(4): 398-400.
- Waza SA, Rastogi NK, Verulkar SB, Sharma D, Bano DA (2014) Microsatellite based estimation of genetic diversity between parental lines and its relationship with yield potential of rice hybrids. *Bioinfolet- A Quart J Life Sci* 11(2): 591-593.
- Xiong L, Wang R, Mao G, Koczan JM (2006) Identification of drought tolerance determinants by genetic analysis of root response to drought stress abscisic acid. *Pl Physiol* 142: 1065-1074.
- Yadav S, Lakshmi N, Singh V, Patil A, Tiwari Y, Nagendran E, Satish P, Maheshwar M, Venkateshwarlu B (2013) *In vitro* screening of *Vigna mungo* to PEG induced moisture deficit stress. *J Agrono Crop Sci* 18: 55–60.
- Zhao T, Dai A (2015) The magnitude and causes of global drought changes in the twenty–first century under a low–moderate emissions scenario. *J Clim* 28: 4490-4512.