Environment and Ecology 41 (4): 2274—2281, October—December 2023 Article DOI: https://doi.org/10.60151/envec/EIUA4469 ISSN 0970-0420

# Evaluation of Bioagents against *Fusarium oxysporum* f. sp. *pisi* Causing Wilt Disease in Pea

### Swarna Kurmi, Sanjeev Kumar, Sanjay Kharte, Rahul Patidar

Received 16 May 2023, Accepted 14 August 2023, Published on 12 October 2023

### ABSTRACT

In this study, in vitro potential of six selected species of Trichoderma, including Trichoderma viride, Trichoderma virens, Trichoderma harzianum, Trichoderma longibrachiatum, Trichoderma ressei and Trichoderma asperellum as well as the of two bacterial isolates Pseudomonas fluorescens and Bacillus subtilis were evaluated against Fusarium oxysporum f. sp. pisi causing wilt of pea. Trichoderma virens recorded maximum percent growth inhibition (81.97%). In volatile compounds percent growth inhibition 86.80 % was recorded highest in Trichoderma harzianum and percent growth inhibition in non-volatile compound will increase as concentration of bioagents increases and 100% growth inhibition was recorded in Trichoderma viride, Trichoderma virens, Trichoderma harzianum and in Trichoderma ressei at 15% concentration.

**Keywords** *Bacillus, Fusarium, Pseudomonas, Trichoderma*, Wilt.

### **INTRODUCTION**

A major cash crop for farmers in Madhya Pradesh is the rabi pea (Pisum sativum). Numerous bacterial, fungal, and viral diseases affect this crop, and under the appropriate condition, diseases can greatly reduce both productivity and quality. The crop production is lower than what is possible due to biotic and abiotic stress. An economically significant disease, Fusarium wilt results in losses of between 30 and 40% (Gupta and Gupta 2019). In plants wilt symptoms are more prominent at 3 to 5 week old plants. Cotyledons droop and wither in early seedlings. Fusarium oxysporum f. sp. *pisi* is soil borne and is capable of saprophytic survival on crop residues in the soil for up to eight years. Because the required large-scale soil application of chemicals is costly, dangerous, and disturbs the biological balance, chemical control of the disease is consequently difficult, impractical, and uneconomical. Hence, efforts have to be made to curtail pathogen activity and restricting losses below economic threshold level by choosing alternative methods. Of late biocontrol methods involving manipulation of antagonistic rhizosphere microflora such as Trichoderma, Pseudomonas or Bacillus against soil borne plant pathogens. Certain species of Bacillus and Pseudomonas can be used as plant growth-promoting bacteria and biological control agents for several plant pathogens, as they are harmless to humans and animals while also being eco-friendly (Rathore et al. 2020). Recently Trichoderma is widely used as seed treatment and soil mix that promote the root growth and protect plant from infection of soil borne fungal

Swarna Kurmi<sup>1</sup>\*, Sanjeev Kumar<sup>2</sup>, Sanjay Kharte<sup>3</sup>, Rahul Patidar<sup>4</sup> <sup>1, 3, 4</sup> PhD Scholar, <sup>2</sup>Assistant Professor

<sup>&</sup>lt;sup>1,2,3</sup>Department of Plant Pathology, College of Agriculture, JNKVV

Jabalpur 482004, MP, India

<sup>&</sup>lt;sup>4</sup>Department of Entomology, College of Agriculture, RVSKVV Gwalior 474001, MP, India

Email : prachikurmi24@gmail.com \*Corresponding author

pathogens. Considering economic importance of disease investigation was carried out to eco-friendly management (*in vitro*) of wilt disease of pea causing *Fusarium oxysporum* f. sp. *pisi*.

### MATERIALS AND METHODS

#### Isolation and purification of the pathogen

Small piece of infected tissues 1-2 mm in size from the propelling edge of the wilted root, with healthy segments were cut with sharp blade, washed well in distilled water to take out dust stuck to the tainted pieces. Pieces were pre-treated 1% NaOC1 (*Sodium hypochlorite*) for 1 minute and consequently washed well in three changes of disinfected refined water. The pieces were then moved to PDA test tube with the help of inoculating needle under aseptic condition and incubated at  $28 \pm 1$ °C. After 72 hrs, fragments of hyphal growth from the growing tips were moved to new PDA test tube. Pure culture was made, following rehashed hyphal tip transfer method.

# Effect of bioagents on growth and sporulation of *Fusarium oxysporum*

For screening of the main antagonist against Fusarium oxysporum dual culture strategy created by Morton and Straube (1955) was taken on. Twenty ml sterilized liquefied PDA medium was filled into sanitized petriplates @ 20 ml/plate aseptically permitted to solidify, then, at that point, then 5 mm discs of the target pathogen and the antagonistic cut with the help of sterilized cork borer were put on PDA around 4 cm separated one another incubated in BOD incubator at  $28 \pm 1^{\circ}$ C. Three replications were kept up with for every treatment. Observation on radial growth of bioagents and test pathogen was recorded after 96 hrs of incubation. Inhibition of mycelial growth and production of spore of target pathogen over check was calculated by following formula (Vincent 1947). In order to study the viability of target pathogen, isolation was done by transferring 5 mm mycelial disc cut by cork borer from the zone where the test fungus was already overgrown by the antagonist on PDA medium.

### Effect of volatile compounds from biocontrol

# agents on radial growth and sporulation of *F. oxysporum*

The effects of volatile compounds from all bioagents on radial growth of *Fusarium oxysporum* were studied as per the method given by Dennis and Webster (1971). The two bottom portion of petriplates containing PDA were inoculated with a 5 mm disc of pathogen and other and sealed with cellophane adhesive tape and incubated in BOD incubator at 28  $\pm$ 1°C. The petriplate containing PDA without antagonist serves as control. The observations on the radial growth of the test fungus were recorded after 120 hrs of incubation at 28  $\pm$  1°C. The radial growth of the test fungus in the treatment in comparison with that of check gave percent growth inhibition.

# Efficacy of non-volatile compounds from biocontrol agents radial growth and sporulation of *F. oxysporum*

The biocontrol agents were grown in *Potato dextrose* broth at 27°C with irregular shaking at 150 rpm. The metabolites were collected after 15 days and filtered. The sterilized filtrate was amended in PDA to make 5, 10 and 15% concentration in petriplates. The solidified agar plates in sets of three were inoculated at the middle with 5 mm diameter mycelial disc of target pathogen and incubated at  $28 \pm 1$ °C for 96 hrs. The plates without filtrate used as control. The colony diameter was recorded and percent inhibition of radial growth was determined utilizing the formula given by Vincent (1947). The statistical method used to conduct the experiments was in Completely Randomized Design method, during 2020-2021.

### **RESULTS AND DISCUSSION**

#### Evaluation of antagonistic efficacy of bioagents

Bioagents, including fungi and bacteria, were tested for their effects on radial growth and sporulation of the test pathogen *Fusarium oxysporum* f. sp. *pisi*. Data on radial growth, percent growth inhibition and sporulation after 124 hrs of incubation are also shown in Table 1 and Fig. 1. The most effective pathogen was *Trichoderma virens* ( $T_2$ ), with the highest rate of growth inhibition (81.97%), *Trichoderma reesei* 

		Du	ual culture		Volatile compounds			
	Treatment	Radial growth of pathogen	% growth inhibition	Sporulation	Radial growth of pathogen	% growth inhibition	Sporulation	
	Trichoderma viride	31.70	54.27	++	26.45	63.45	++	
2	Trichoderma virens	12.50	81.97		20.31	71.93	+	
2	Trichoderma harzianum	29.60	57.30	—	9.55	86.80	-	
1	Pseudomonas fluorescens	38.40	44.60	_ ++	31.16	56.95	+	
5	Bacillus subtilis	45.20	34.79	++	30.88	57.34	+	
	Trichoderma longibrachiatum	30.10	56.57	-	31.73	56.15	+	
,	Trichoderma ressei	25.20	63.64		27.78	61.61	+	
, o	Trichoderma asperellum	31.22	54.96	_	34.00	53.03	++	
Γ <sub>9</sub> °	Control	69.31	0.00	—	72.37	0.00	++++	
	SE (m)	0.62			0.68			
	CD (0.5)	1.86			2.05			

Table 1. Antagonistic efficacy of bioagents under dual culture technique and their volatile compounds against Fusarium oxysporum f. sp. pisi after 124 hrs of incubation.

(T<sub>7</sub> - 63.64%), *Trichoderma longibrachiatum* (T<sub>6</sub> - 56.57%), *Trichoderma halzianum* (T<sub>3</sub> - 57.30%). The least effective pathogen was *Bacillus subtilis* 

 $(T_5)$ , inhibited by only 34.79%, followed by *Pseudomonas fluorescens* (T<sub>4</sub>-44.60%) and *Trichoderma viride* (T<sub>1</sub> - 54.27%). Among them, sporulation was



Fig. 1. Antagonistic efficacy of bioagents against Fusarium oxysporum by using dual culture technique.



Pseudomonas fluorescens

Bacillus subtilis

Control

Fig. 2. Antagonistic efficacy of bioagents against Fusarium oxysporum f. sp. pisi by using volatile compound.

well observed in his Trichoderma viride, Bacillus subtilis and Pseudomonas fluorescens bacteria. No sporulation was recorded with the other treatments.

## Evaluation of antagonistic efficacy of volatile and non-volatile compounds of bioagents

Bioagents were also evaluated for their effect on test pathogen as they produce some volatile compounds during primary and secondary metabolism having tendency to vaporize which help in significantly reduce radial growth, percent growth inhibition and sporulation in in vitro and data on all presented in Table 1 and Fig. 2. Percent growth inhibition 86.80% was recorded highest in Trichoderma harzianum followed by Trichoderma virens (T2- 71.93%) and Trichoderma viride (T<sub>1</sub>-63.45%). Lowest was recorded in Trichoderma asperellum (T<sub>8</sub>-53.03%) followed by Trichoderma longibrachiatum (T<sub>6</sub>- 56.15%), and Pseudomonas fluorescens (T<sub>4</sub>- 56.95%). Almost all treated plates results sporulation except Trichoderma harzianum there was no sporulation recorded. Excellent sporulation was recorded in control. Trichoderma asperellum and Trichoderma viride result good sporulation and poor sporulation was recorded in the remaining treatments.

Non-volatile compounds extracted from the liquid culture were assessed @ 5, 10 and 15% for their antifungal activity on radial growth, percent growth inhibition and sporulation of test pathogen and data presented in Table 2 and Fig. 3. Percent growth inhibition was recorded highest in Trichoderma virens (T<sub>2</sub>- 82.01%), followed by Trichoderma viride (T<sub>1</sub>-64.74%), Trichoderma harzianum ( $T_3$ - 63.54%) and Trichoderma ressei (T<sub>7</sub>-63.54%). Lowest inhibition was recorded in Bacillus subtilis (T<sub>5</sub>-38.20%), followed by Pseudomonas fluorescens (40.76%), Trich-



Fig. 3. Effect of non-volatile compounds (15%) of bioagents against Fusarium oxysporum.

oderma longibrachiatum (56.83%) and Trichoderma asperellum (62.47%). Sporulation was excellent in control plate. Good sporulation was recorded in Pseudomonas fluorescens. No sporulation was recorded in Trichoderma virens, Trichoderma viride and Trichoderma harzianum treated plate and remaining four show poor sporulation. At 10% growth inhibition was recorded highest in Trichoderma virens (T<sub>2</sub>-92.57%), followed by Trichoderma viride (T<sub>1</sub>-88.54%), Trichoderma harzianum (T<sub>3</sub>- 85.15%) and Trichoderma ressei (T<sub>7</sub>-73.90%). Lowest inhibition was recorded in Bacillus subtilis (T<sub>5</sub>-51.10%), followed by Pseudomonas fluorescens (57.79%), Trichoderma longibrachiatum (70.17%) and Trichoderma asperellum (71.54%). Sporulation was excellent in control plate followed by Bacillus subtilis. Good sporulation was recorded in Pseudomonas fluorescens and Trichoderma longibrachiatum. No sporulation was recorded in Trichoderma virens treated plate and remaining four show poor sporulation ; and at 15% all tested bioagents were found effective and percent growth inhibition was recorded lowest in *Bacillus subtilis* ( $T_5$ -85.5%), followed by *Pseudomonas fluorescens* (87.00%), *Trichoderma asperellum* (89.0%) and *Trichoderma longibrachiatum* (93.6%). Sporulation was excellent in only control plate. No sporulation was recorded in any treatment except *Trichoderma asperellum* show poor sporulation.

### DISCUSSION

As a potent and suitable biological control agent for managing soil-borne diseases, *Trichoderma* has grown in favor. *Trichoderma* species have shown potential against a number of significant soil-borne diseases, including *Pythium, Fusarium, Rhizoctonia, Sclerotium* and *Macrophomina* (Choudhary and Mohanka 2012, Kumar *et al.* 2012, Ommati and Zaker

Table 2. Effect of non-volatile compounds (5, 10 and 15%) of bioagents against *F. oxysporum* f. sp. *pisi* under *in vitro* of condition after 124 hrs of incubation.

-				5%			10%		15%		
	Treatment	Radial growth of path- ogen	% growth inhibition	Sporula- tion	Radial gro- wth of path- ogen	% growth inhibi- tion	Sporula- tion	Radial growth of pathogen	% growth inhibition	Sporula- tion	
Τ,	Trichoderma viride	24.60	64.74	+	8.11	88.54	-	0.00	100.0	-	
T <sub>2</sub>	Trichoderma virens	12.55	82.01	-	5.26	92.57	-	0.00	100.0	-	
Τ <sub>3</sub>	Trichoderma harzi- anum	25.44	63.54	+	10.51	85.15	-	0.00	100.0	-	
Τ 4	Pseudomonas fluo-										
	rescens	41.33	40.76	++	29.87	57.79	++	10.12	87.0	-	
T 5 T 6	Bacillus subtilis Trichoderma longibra-	43.12	38.20	+++	34.61	51.10	+	11.31	85.5	-	
	chiatum	30.12	56.83	++	21.11	70.17	+	5.02	93.6	-	
T <sub>7</sub>	Trichoderma ressei	25.44	63.54	+	18.47	73.90	+	0.00	100.0	-	
T <sub>s</sub>	Trichoderma aspere-										
0	llum	26.19	62.47		20.14	71.54	+	8.61	89.0	+	
T <sub>9</sub>	Control	69.77	0.00		70.77	0.00	++++	78.01	0.0	++++	
	SE (m)	0.67			0.51			0.43			
	CD (0.5)	2.01			1.54			1.29			

2012, Raut *et al.* 2014, Magar *et al.* 2020). Each of the investigated bioagents differentially restricted the pathogen's growth and increased the pathogen colony in the dual culture test as compared to the control. *Trichoderma virens* (81.97%) showed the greatest *Fusarium oxysporum* f. sp. *pisi* inhibition in solo culture, followed by *Trichoderma ressei* and *Trichoderma harzianum*. This result is in partial consonance with Kumar *et al.* (2009), Hamid *et al.* (2012), Kim and Knudsen 2013, Patel and Patel 2014, Kumar *et al.* (2016), Anuragi and Sharma (2016), Hassan *et al.* (2013).

By observing their radial growth and sporulation, the volatile compounds from the bioagents *Trich*oderma viride, *Trichoderma virens*, *Trichoderma* harzianum, *Trichoderma longibrachiatum*, *Tricho*derma ressei, *Trichoderma asperellum*, *Pseudomo*nas fluorescens and Bacillus subtilis were assessed against Fusarium oxysporum f. sp. pisi. Trichoderma harzianum had the highest percent growth inhibition, followed by *Trichoderma virens* and *Trichoderma* viride. which conflicts with research by other researchers, including Singh et al. (2002), Kumar et al. (2009), Hamid et al. (2012), Kapoor et al. (2012), Kumar et al. (2014), Nagamani et al. (2016), Singh et al. (2016) and Cherkupally et al. (2017). Similar to this, the non-volatile components from bioagents were tested against *Fusarium oxysporum* f. sp. *pisi* at 5, 10, and 15% concentrations. The non-volatile substance from each individual *Trichoderma* treatment showed growth suppression of the test pathogen. Maximum *Fusarium oxysporum* f. sp. *pisi* growth and sporulation suppression was seen in *Trichoderma virens, Trichoderma harzianum* and *Trichoderma virens, Trichoderma harzianum* and *Trichoderma ressei* during single treatment. Many researchers, including Kumar and Dabbas(2016), Dubey *et al.* (2007) and Chaudhary *et al.* (2017), also obtained similar results. These outcomes provide strong support for the current conclusions.

Trichoderma harzianum SQR-T037, a new strain isolated by Raza et al. (2013), is tested in vitro for its ability to inhibit Fusarium oxysporum using both its volatile and non-volatile chemicals. To examine the antifungal effects of the volatile and nonvolatile metabolites generated by the Trichoderma T36 and T50 strains. A study was conducted by Raut and coworkers (2014) on Fusarium graminearum, Rhizoctonia solani and Pythium ultimum and discovered that the volatile component of Trichoderma T36 resulted in stronger inhibition than T 50 against all the pathogens. Trichoderma virens, Trichoderma viride and Trichoderma harzianum were the targets of Li et al. (2018) research, which revealed that these bioagents had the potential to inhibit the growth of several Fusarium oxysporum strains. They also draw the conclusion that all species of Trichoderma can detect the volatiles produced by Fusarium spp. and can distinguish their presence by sensing them. Trichoderma then gets ready to combat pathogens by producing secondary metabolite compounds or volatile compounds. Trichoderma asperellum GDSF 1009 (CGMCC NO. 9512), Trichoderma asperelloides Z4-1 (CGMCC NO. 40245), Trichoderma harzianum 10569 (CG-MCC NO. 40246), and T. asperellum 10264 (CGMCC NO. 22404) were combined to determine the best consortium for co-culture, the factors underlying the levels of Fusarium oxysporum antagonistic and growth-promoting activity in plants, as well as the enhancement of seed germination in monocultures of a single strain (Hao et al.2022).

### CONCLUSION

Six spp. of *Trichoderma*, including *T. viride*, *T. virens*, *T. harzianum*, *Trichoderma longibrachiatum*, *T. ressei* and *T. asperellum* and two bacterial isolates *Pseudomonas fluorescens* and *Bacillus subtilis* were evaluated for their volatile and non-volatile compounds, *in vitro* potential. All tested bioagents were effective against *Fusarium* in various degree. In dual culture *Trichoderma virens* found effective, in volatile compounds 86.80 % growth inhibition was recorded in *Trichoderma harzianum*, In non volatile compound, 100% growth inhibition was recorded in *Trichoderma virens*, *Trichoderma harzianum* and in *Trichoderma ressei* at 15% concentration.

### REFERENCES

- Anuragi M, Sharma TK (2016) Biocontrol of chickpea wilt disease by *Fusarium oxysporum* f. sp. *ciceris* with rhizosphere mycoflora. *Flora and Fauna* 22 (2): 201–209.
- Chaudhary A, Sharma H, Jehani M, Sharma JK (2017) Seed mycoflora associated with pigeonpea [*Cajanus cajan* (L.) Millsp.], their significance and the management. *J Pure Appl Microbiol* 11 (1): 567—575.
- Cherkupally R, Amballa H, Reddy BN (2017) In vitro antagonistic activity of Trichoderma species against Fusarium oxysporum f. sp. melongenae. Int J Appl Agricult Res 12 (1): 87—95.
- Choudhary S, Mohanka R (2012) In vitro antagonism of indigen-

ous trichoderma isolates against phytopathogen causing wilt of lentil. *Adv Res in Pharmaceut Biologi* 2 (3) : 195–202.

- Dennis C, Webster J (1971) Antagonistic properties of speciesgroups of *Trichoderma*: I. Production of non-volatile an tibiotics. *Transac British Mycological Soc* 57: 25—39.
- Dubey SC, Suresh M, Singh B (2007) Evaluation of *Trichoderma* species against *Fusarium oxysporum* f. sp. *ciceris* for integrated management of chickpea wilt. *Biologi Control* 40 (1): 118—127.
- Gupta SK, Gupta M (2019) *Fusarium* wilt of pea-A mini review. *Pl Dis Res* 161 (1) : 1—9.
- Hamid A, Bhat NA, Sofi TA, Bhat KA, Asif M (2012) Management of root rot of pea (*Pisum sativum* L.) through bioagents. *Afr J Microbiol Res* 6 (44) : 7156—7161.
- Hao D, Lang B, Wang Y, Wang X, Liu T, Chen J (2022) Designing synthetic consortia of *Trichoderma* strains that improve antagonistic activities against pathogens and cucumber seedling growth. *Microbial Cell Factories* 21 : 234.
- Hassan SAE, Gowen SR, Pembroke P (2013) Use of *Trichoderma* hamatum for biocontrol of lentil vascular wilt disease : Efficacy, mechanisms of interaction and future prospects. J Pl Prot Res 53 (1) : 12–25.
- Kapoor S, Jaiswal A, Shukla DN (2012) Eco- friendly strategies for management of *Fusarium* wilt of *Pisum sativum* L. Afr J Microbiol Res 6 (48): 7397–7400.
- Kim TG, Knudsen GR (2013) Relationship between the biocontrol fungus *Trichoderma harzianum* and the phytopathogenic fungus *Fusarium solani* f. sp. *pisi. Appl Soil Ecolog* 68 : 57—60.
- Kumar S, Dabbas MR (2016) Evaluation of different botanicals (leaf extract) management of *Fusarium wilt* of vegetable pea. *Ind J Agricult Appl Sci* 2 : 4.
- Kumar S, Dabbas MR, Mishra O, Singh S (2016) Effect of chemical management against wilt disease of vegetablepea caused by *Fusarium oxysporum* f. sp. *pisi. J Mycopathol Res* 54 (1): 77—80.
- Kumar S, Singh J, Biswas SK, Naresh P, Dabas MR (2012) Effect of culture media, temperature, pH and host range on the growth of *Fusarium oxysporum* f.sp. *pisi*. J Mycopathol Res 50 (1): 73—76
- Kumar S, Manibhushan Thakur M, Rani A (2014) *Trichoderma* : Mass production, formulation, quality control, delivery and its scope in commercialization in India for the management of plant diseases. *Afr J Agricult Res* 9 (53) : 3838–3852.
- Kumar S, Upadhyay JP, Rani A (2009) Evaluation of *Trichoderma* spp. against *Fusarium udum* Butler causing wilt pea. *J Biological Control* 23 (3) : 329–332.
- Li Ningxiao, Alsayed A, Wenzhao W, Md Islam, Khoshnood N, Xingzhong Liu, Seogchan K (2018) Volatile Compound-Mediated Recognition and Inhibition Between *Trichoderma* biocontrol Agents and *Fusarium oxysporum*. Original research.
- Magar SJ, Patange AS, Somwanshi SD (2020) In vitro efficacy of fungicides, bioagents and silver nanoparticles against Fusarium oxysporum f. sp. ciceri. Ind Phytopathol 73 : 65—69.
- Morton DJ, Stroube WH (1955) Antagonistic and stimulating effects of soil micro-organism of Sclerotium. *Phytopathol* 45: 417–420.
- Nagamani P, Biswas MK, Bhagat S (2016) Efficacy of native *Trichoderma* spp. to the management of soil borne pathogens

in chickpea (*Cicer arietinum* L International Conference "Plant, Pathogens and People" February 23-27. New Delhi, India.

- Ommati F, Zaker M (2012) Evaluation of some *Trichoderma* isolates for biological control of potato wilt disease (*Fusarium oxysporum*) under laboratory and greenhouse conditions. *J Crop Prot* 1 (4) : 279–286.
- Patel R, Patel D (2014) Screening of *Trichoderma* and antagononistic analysis of a potential strain of *Trichoderma* for production of a bioformulation. *Int J Sci Res Publ* 4 (10) : 16.
- Rathore R, Vakharia DN, Rathore DS (2020) *In vitro* screening of different *Pseudomonas fluorescens* isolates to study lytic enzyme production and growth inhibition during antagonism of *Fusarium oxysporum* f. sp. *cumini*, wilt causing pathogen of cumin. *Egypt J Biol Pest Cont* 30 : 57.
- Raut I, Calin M, Vasilescu G, Doni MB, Tatiana S, Jecu L (2014)

Effect of non-volatile compounds of *Trichoderma* spp. against *Fusarium graminearum*, *Rhizoctonia solani* and *Pythium ultimum*. Sci Bull Series F. Biotechnologies XVII.

- Raza W, Faheem F, Yousaf S, Rajer FU, Yameen M (2013) Volatile and non-volatile antifungal compounds produced by *Trichoderma harzianum* SQR-T037 suppressed the growth of *Fusarium oxysporum* f. sp. *niveum. Sci Letters* 21—24.
- Singh R, Kumar R, Sharma A (2002) Evaluation of some pea lines for resistant to wilt. *J Mycol Pl Pathol* 2 : 150—151.
- Singh R, Laxmikant, Singh M (2016) Management of root rot of pea (*Pisum sativum* L.) through commercially available bioagent formulation and fungicide. 6<sup>th</sup> International Conference "Plant, Pathogens and People" February 23-27, New Delhi, India.
- Vincent JM (1947) Distortion of fungal hyphae in presence of certain inhibitors. *Nature* 154 : 158.