Environment and Ecology 41 (3C): 1953—1959, July—September 2023 Article DOI: https://doi.org/10.60151/envec/WYKC4262 ISSN 0970-0420

# **Optimizing Priming Concentration and Duration of PEG 6000 for Improving Seed Germination and Vigour in Chickpea (***Cicer arietinum* L.)

# Anish Choudhury, Sanjoy Kumar Bordolui, Jui Ray

Received 15 March 2023, Accepted 21 June 2023, Published on 4 September 2023

### ABSTRACT

This study was undertaken with different concentration and duration of PEG-6000 to observe the effect of germination and seedling vigour in chickpea seeds. The experiment was conducted in seed testing laboratory, Department of Seed Science and Technology, Bidhan Chandra Krishi Viswavidyalaya, Mohanpur, Nadia, West Bengal, India during 2020-2021 following Complete Randomized Block Design with three replications in order to standardize the appropriate concentration and duration while soaking the seeds within PEG-6000 solution for screening a range of duration concentrations viz, 10 g liter<sup>-1</sup> for 6 hrs, 10 g liter<sup>-1</sup> for 8 hrs, 10 g liter<sup>-1</sup> for 10 hrs, 15 g liter<sup>-1</sup> for 6 hrs, 15 g liter<sup>-1</sup> for 8 hrs, 15 g liter<sup>-1</sup> for 10 hrs, 20

<sup>1, 3</sup> Research Scholar, <sup>2</sup> Assistant Professor

Department of Seed Science and Technology, Bidhan Chandra Krishi Viswavidyalaya, Mohanpur, Nadia, West Bengal 741252, India

Email : sanjoy\_bordolui@rediffmail.com \*Corresponding author

g liter<sup>1</sup> for 6 hrs, 20 g liter<sup>1</sup> for 8 hrs, 20 g liter<sup>1</sup> for 10 hrs, 25 g liter<sup>-1</sup> for 6 hrs, 25 g liter<sup>-1</sup> for 8 hrs, 25 g liter<sup>-1</sup> for 10 hrs. Untreated dry seeds were considered as control. It was found that all the priming treatments showed significance difference in overall including the control. The best performer or the better result was shown by T<sub>8</sub> which is 20 gm/ liter solution with the soaking duration of 8 hrs. It was highest performer in germination percentage (93.77), root length (6.57 cm), shoot length (16.57 cm), seedling fresh weight (1.66 g), seedling dry weight (0.14 g), seedling vigour index I (2137.07), time to 50% germination (2.33) days) and germination index (77.82). This study showed that seed priming with osmotic solution could improve some seedling parameters in chickpea. In seed priming, its simplicity such as no requirements for extensive equipment and chemicals could be used method for overcoming problems related to a poor germination and further seedling establishment and helps in sustaining agriculture with a cost effective approach and with the help of seed priming treatments which are non-toxic and from eco-friendly sources.

**Keywords** Chickpea, Germination, PEG 6000, Osmo priming, Vigour.

### **INTRODUCTION**

Among the pulses chickpea is the highly nutritious crop. By consumption of crops, it will provide the nutritional security of people as it is a good source of protein, carbohydrate and several essential minerals.

Anish Choudhury<sup>1</sup>, Sanjoy Kumar Bordolui<sup>2\*</sup>, Jui Ray<sup>3</sup>

Chickpea (Cicer arietinum L.) is a self-pollinated crop. It is diploid (2n=16) species with genome size 1C=740 Mbp (Arumuganathan and Earle 1991) and belonging to sub family Papilionaceae of the family Leguminaceae (Poehlman and Sleper 1995). It is the world's third-most important food legume (pulse) and is consumed as a high-quality protein food with India being the world's largest producer and consumer of pulses. Chickpea contains 17-22 % protein and 60-64% carbohydrates and helps in sustaining the productivity of the cropping systems through their ability to fix atmospheric nitrogen. India is the largest chickpea producing country accounting for 64% of the global chickpea production. Moreover, in rice-based cropping systems and rainfed regions, chickpea planting is often delayed due to late harvest of rice and/or untimely rainfall. This delay in chickpea planting is often tied with low temperature (Srinivasan et al. 1998), which limits the germination and stand establishment (Jame and Cutforth 2004). Priming of seeds in osmoticums such as mannitol, polyethylene glycol and sodium chloride has been reported to be an economical, simple and a safe technique for increasing the capacity of seeds to osmotic adjustment and enhancing seedling establishment and crop production under stressed conditions (Kaur et al. 2001). This could be due to faster emergence of roots and shoots, lower incidence of resowing, more vigorous plants, better drought tolerance, earlier flowering, earlier harvest and higher grain yield under adverse conditions (Amzallag et al. 1990, Cayuela et al. 1996). The beneficial effect of priming on germination has been reported in tomato seeds (Ozbingol et al. 1998). Saglam et al. (2010) reported priming treatments decreased the effects of water stress and (Ghassemi-Golezani et al. 2013) observed that osmo and hydropriming treatments to lentil seeds increased the plant height, number of pods and number of seeds per plant, the 1,000 grain weight, the biological yield, the grain yield and the harvest index when compared to the control. The aim of this study was to observe the effects of osmo-priming treatments with different concentration and duration to chickpea seeds, to analyze the influence of such a treatment over seed germination properties and identify the best concentration with duration for the betterment of plant characteristics, yield components in an experiment conducted with field conditions.

### MATERIALS AND METHODS

The experiment was carried out in seed testing laboratory, Department of Seed Science and Technology, Bidhan Chandra Krishi Viswavidyalaya, Mohanpur, Nadia, West Bengal, India during 2021 following Complete Randomized Design with three replications. Chickpea (vatiety-Anuradha) was collected from AICRP pulses in Bidhan Chandra Krishi Viswavidyalaya, Mohanpur, Nadia, West Bengal for this investigation. Seed priming was done with 10 g liter<sup>1</sup> for 6 hrs (T<sub>1</sub>), 10 g liter for 8 hrs (T<sub>2</sub>), 10 g liter<sup>-1</sup> for 10 hrs (T<sub>3</sub>), 15 g liter<sup>1</sup> for 6 hrs (T<sub>4</sub>), 15 g liter<sup>1</sup> for 8 hrs (T<sub>5</sub>), 15 g liter<sup>1</sup> for 10 hrs (T<sub>6</sub>), 20 g liter<sup>1</sup> for 6 hrs  $(T_2)$ , 20 g liter<sup>-1</sup> for 8 hrs  $(T_2)$ , 20 g liter<sup>-1</sup> for 10 hrs ( $T_9$ ), 25 g liter<sup>-1</sup> for 6 hrs ( $T_{10}$ ), 25 g liter<sup>-1</sup> for 8 hrs  $(T_{11})$ , 25 g liter<sup>-1</sup> for 10 hrs  $(T_{12})$ . Non-primed seeds were the control  $(T_0)$ . Following observations were taken for this experiment.

# Time to 50% germination

According to Association of Official Seed Analysis 1983 number of seeds germinated was recorded in daily basis. The time to obtain 50% germination ( $T_{50}$ ) was calculated according to the following formulae given by Coolbear *et al.* (1984) which was modified by Farooq *et al.* (2005).

$$\mathbf{T}_{50} = \mathbf{t}_{i} + \frac{\left(\begin{array}{c} \frac{\mathbf{N}}{2} - \mathbf{n}_{i} \right) \left(\mathbf{t}_{j} - \mathbf{t}_{i} \right)}{(\mathbf{n}_{i} - \mathbf{n}_{i})}$$

Where, N stands for final number of germination and  $n_i$ ,  $n_j$  are cumulative number of seeds germinated by adjacent counts at times  $t_i$  and  $t_j$  when  $n_i < N/2 < n_i$ .

### Mean germination time (MGT)

Mean Germination Time (MGT) was calculated with the following equation suggested by Ellis and Roberts (1981).

$$MGT = \frac{\sum D_n}{\sum n}$$

Where D is the number of days counted from the beginning of germination and n indicates the number

of seeds germinated on day D.

### **Germination percentage**

Germination percentage (G) was expressed in percentage. It was calculated as :

 $\frac{\text{Number of normal seedlings produced}}{\text{Total number of seeds used}} \times 100$ 

Where, X is the number of normal seedlings produced and Y denotes total number of seeds taken for germination (ISTA 1996).

# Germination index (GI)

Germination index (GI) was calculated as described in the Association of Official Seed Analysts (AOSA 1990) as the following formulae :

	No. of germi-		No. of germi-	
CI –	nated seeds	т т	nated seeds	
01	Day of first		Day of last	
	count		count	

# Germination energy

According to Ruan *et al.* (2002) energy of germination (GE) should be recorded at 4<sup>th</sup> day after planting. It is the percentage of germinating seeds 4 days after planting relative to the total number of seeds tested.

# Seedling parameters

Root lengths and shoot lengths of ten seedlings were measured at 8 days after germination by glass plate method in the laboratory with the help of a scale and graph paper and average was made out, expressed in centimeter (cm). Fresh weight of ten seedlings was measured with the help of a digital balance. Then seedlings were dried at 60-70°C for two hours in hot air oven and weighed in a digital balance. Both seedling fresh weight and dry weight are expressed in gram (g).

### Vigour index

Vigour index (VI) was calculated by using the for-

Where, 'L' denotes average seedling length (cm) and 'G' indicates germination percentage.

# **RESULTS AND DISCUSSION**

### **Germination index**

Germination index was varied significantly and after counting daily basis of seedling emergence up to eight days, lowest Germination index was observed in control (23.06) preceded by  $T_{4^3}$   $T_1$  and  $T_{12}$ . While,  $T_8$  (43.10) showed the highest germination index followed by  $T_{11}$ ,  $T_5$  and  $T_9$ . But, non-significant difference was observed in between  $T_3$  and  $T_{12}$ ,  $T_3$  and  $T_7$ ,  $T_2$  and  $T_{10}$  (Table 1). Similar type of result was observed by Ray and Bordolui (2022b).

# Root length (cm)

During different duration and concentration root length of seedling was varied significantly and after

**Table 1.** Effect of priming on germination index, root length, shoot length, fresh weight and dry weight of chickpea. Note:  $T_0 = Control$ ,  $T_1=10$  g liter<sup>-1</sup> for 6 hrs,  $T_2=10$  g liter<sup>-1</sup> for 8 hrs,  $T_3=10$  g liter<sup>-1</sup> for 10 hrs,  $T_4=15$  g liter<sup>-1</sup> for 6 hrs,  $T_5=15$  g liter<sup>-1</sup> for 8 hrs,  $T_6=15$  g liter<sup>-1</sup> for 10 hrs,  $T_7=20$  g liter<sup>-1</sup> for 6 hrs,  $T_8=20$  g liter<sup>-1</sup> for 8 hrs,  $T_{12}=25$  g liter<sup>-1</sup> for 6 hrs,  $T_{11}=$  g liter<sup>-1</sup> for 8 hrs,  $T_{12}=25$  g liter<sup>-1</sup> for 10 hrs.

Treat- ment	Germi- nation index	Root length (cm)	Shoot length (cm)	Fresh weight (g)	Dry weight (g)
$ \frac{T_{0}}{T_{1}} T_{2}^{2} T_{3}^{3} T_{4}^{4} T_{5}^{5} T_{6}^{7} T_{7}^{7} T_{8}^{7} T_{9}^{9} T_{10}^{10} T_{11}^{11} T_{12}^{12} SEm (\pm) $	23.06 27.10 30.40 27.97 26.42 36.30 29.07 28.12 43.10 31.47 30.06 42.40 27.76 <b>0.14</b>	4.00 4.70 4.50 4.52 4.97 6.11 4.52 5.44 6.57 4.16 2.67 3.46 5.40 <b>0.154</b>	7.56 9.56 12.55 13.93 13.28 14.79 14.58 15.56 16.57 13.29 12.45 12.81 12.35 <b>0.142</b>	0.91 0.85 0.95 1.07 1.24 1.55 1.08 1.46 1.66 1.44 1.10 1.12 1.38 <b>0.038</b>	0.09 0.10 0.08 0.11 0.13 0.14 0.09 0.12 0.14 0.13 0.09 0.07 0.13 0.009
LSD (0.05)	0.41	0.449	0.414	0.112	0.027

Treatment	Germination percentage (transform value)	Vigor index	Time to 50% germination (days)	Mean germi- nation time (days)	Germination energy (%)
T	85.83 (67.86)	992.18	3.55	4.40	51.57 (45.88)
Τ <sub>1</sub>	89.80 (71.37)	1265.44	2.74	3.83	65.64 (54.09)
T,	88.60 (70.26)	1660.46	2.40	3.53	76.78 (61.18)
T,	87.90 (69.64)	1609.60	2.72	3.33	65.49 (54.00)
T,	87.87 (69.61)	1601.85	2.59	3.89	65.20 (53.83)
T,	91.53 (73.08)	1553.85	2.53	3.69	71.66 (57.82)
T <sub>6</sub>	87.37 (69.18)	1844.48	2.88	3.85	59.67 (50.56)
T <sub>2</sub>	87.70 (69.46)	1858.32	2.72	3.42	67.46 (55.20)
T.	93.77 (75.56)	2137.07	2.33	3.34	77.82 (61.89)
T <sub>0</sub>	93.60 (75.35)	1611.46	2.43	3.56	71.59 (57.77)
T_10	87.27 (69.08)	1295.91	2.50	3.25	69.47 (56.44)
T	93.63 (75.38)	1515.20	2.95	3.05	71.97 (58.02)
T <sub>12</sub>	90.93 (72.48)	1611.66	2.96	4.04	59.61 (50.52)
SËm (±)	0.700	28.650	0.086	0.129	0.422
LSD (0.05)	2.046	83.743	0.250	0.376	1.235

**Table 2.** Effect of priming on germination percentage, vigor index, Time to 50% germination, Mean germination time and germination energy of chickpea. Note :  $T_0 = Control$ ,  $T_1=10$  g liter<sup>1</sup> for 6 hrs,  $T_2=10$  g liter<sup>1</sup> for 8 hrs,  $T_3=10$  g liter<sup>1</sup> for 10 hrs,  $T_4=15$  g liter<sup>1</sup> for 6 hrs,  $T_5=15$  g liter<sup>1</sup> for 8 hrs,  $T_6=15$  g liter<sup>1</sup> for 10 hrs,  $T_7=20$  g liter<sup>1</sup> for 6 hrs,  $T_8=20$  g liter<sup>1</sup> for 8 hrs,  $T_9=20$  g liter<sup>1</sup> for 10 hrs,  $T_{10}=25$  g liter<sup>1</sup> for 6 hrs,  $T_{11}=$  g liter<sup>1</sup> for 8 hrs,  $T_{12}=25$  g liter<sup>1</sup> for 10 hrs.

eight days of final count maximum seedling root length was observed for  $T_8$  (6.57 cm) followed by  $T_5$ and  $T_7$  respectively, while it was minimum for  $T_{10}$ (2.67 cm) (Table 1). Although  $T_1$ ,  $T_2$ ,  $T_3$  and  $T_6$ ,  $T_3$ and  $T_4$ ,  $T_4$  and  $T_{12}$ ,  $T_0$  and  $T_9$ ,  $T_7$  and  $T_{12}$ ,  $T_2$ ,  $T_3$ ,  $T_6$  and  $T_9$  showed non-significant difference among themselves (Table 1). Root and shoot lengths increased in seeds due to priming as compared to non-primed seeds reported by Kaur *et al.* (2000), Choudhury and Bordolui (2022a).

### Shoot length (cm)

Significant variation was observed for seedling shoot length, when average was taken over treatments; Considering shoot length, the longest seedling shoot length was recorded for  $T_8$  (14.57 cm) followed by  $T_7$  and  $T_5$  while shortest shoot length was observed in Control (7.56 cm) preceded by  $T_1$  and  $T_{12}$ . Significant difference was noted for shoot length in overall though non-significant difference was observed in between  $T_2$ ,  $T_{10}$  and  $T_{12}$ ,  $T_2$ ,  $T_{10}$  and  $T_{11}$ ,  $T_4$  and  $T_9$ ,  $T_6$  and  $T_8$  (Table 1). The result corroborates the findings of Farooq *et al.* (2008), Choudhury and Bordolui (2022b).

# Fresh weight (g)

Fresh weight of five seedlings was recorded ant it was significantly difference varied in fresh weight after PEG 6000 solution. Highest seedling fresh weight was observed for  $T_8$  (1.66 g) followed by  $T_5$  and  $T_7$  while lowest was noted in  $T_1$  (0.85 g) preceded by  $T_0$  and  $T_2$  respectively. But non-significant difference was noticed in between  $T_5$  and  $T_8$ ,  $T_3$ ,  $T_6$ ,  $T_{10}$  and  $T_{11}$ ,  $T_5$ ,  $T_7$  and  $T_9$ ,  $T_5$  and  $T_8$ ,  $T_7$ ,  $T_9$  and  $T_{12}$ ,  $T_0$ ,  $T_1$  and  $T_2$  (Table 1).

# Dry weight (g)

In case of dry weight, it was significantly varied due to priming with different duration and concentration of PEG. Maximum seedling dry weight was noticed for  $T_5$  (0.14 g) followed by  $T_9$  and  $T_7$  respectively while minimum was noticed for  $T_{11}$  (0.07 g) preceded by  $T_2$  and  $T_6$  respectively. Although non-significant difference was observed in between  $T_0$ ,  $T_1$  and  $T_2$ ,  $T_0$ ,  $T_1$  and  $T_3$ ,  $T_3$ ,  $T_4$ ,  $T_7$ ,  $T_9$  and  $T_{12}$ ,  $T_2$ ,  $T_6$ ,  $T_{10}$  and  $T_{11}$ ,  $T_4$ ,  $T_5$ ,  $T_7$ ,  $T_8$ ,  $T_9$  and  $T_{12}$  (Table 1). Mohammadi (2009) reported similar kind of experiment in soybean (*Glycine max* L.).

## Germination percentage

Significant difference was observed in germination percentage. Among the priming treatments, with



Fig 1. Evaluation of seedling vigour under laboratory condition.

# different duration and concentration of PEG 6000, $T_8$ (93.77 %) recorded highest germination percentage followed by $T_{11}$ and $T_9$ . While lowest germination percentage was recorded for $T_0$ (85.83 %) preceded by $T_4$ , and $T_3$ respectively. But, non-significant difference was observed in between $T_1$ , $T_2$ , $T_3$ and $T_4$ , $T_2$ , $T_3$ , $T_4$ , $T_6$ , $T_7$ and $T_{10}$ , $T_0$ , $T_4$ , $T_6$ , $T_7$ and $T_{10}$ , $T_1$ and $T_8$ (Table 2). This result is in agreement with Singh and Rao (1993).

# Vigour index

Considering vigour index, it was varied significantly when average was made over the treatments. Maximum value was calculated for  $T_8$  (2013.08) followed by  $T_1$  and  $T_8$  respectively, minimum vigour index was noted for  $T_0$  (992.18) preceded by  $T_{10}$  and  $T_3$ . Although, vigour index was significantly varied, but some non-significant difference was also noticed in between  $T_1$  and  $T_5$ ,  $T_3$ ,  $T_7$ ,  $T_{10}$  and  $T_{12}$ ,  $T_2$ ,  $T_4$  and  $T_9$ ,  $T_6$ ,  $T_8$  and  $T_{11}$  (Table 2, Fig. 1). Iqbal (2015) conducted similar kind of experiment in cowpea (*Vigna unguiculata* L.), Chakraborty and Bordolui (2021) observed similar kind of experiment in green gram.

# Time to 50% germination (days)

Significant responses were noticed in the priming treatment with different duration and concentration of PEG-6000 solution under laboratory condition. Minimum time to 50% germination was recorded in  $T_8$  (2.33 days) followed by  $T_2$  and  $T_9$ . While, maximum time to 50% germination was observed for  $T_0$  (3.55 days) preceded by  $T_{12}$  and  $T_{11}$ . Although, time to 50% germination was significantly varied, but some non-significant difference was also noticed in between  $T_1$ ,  $T_3$ ,  $T_4$ ,  $T_5$ ,  $T_7$  and  $T_{10}$ ,  $T_2$ ,  $T_4$ ,  $T_5$ ,  $T_9$  and  $T_{10}$ ,  $T_2$ ,  $T_5$ ,  $T_8$ ,  $T_9$  and  $T_{10}$  (Table 2). The result corroborates the findings of Dezfuli *et al.* (2008).

# Mean germination time (days)

Considering mean germination time  $T_{11}$  (3.05 days) had the shortest mean germination time preceded by  $T_8$  and  $T_2$ . While maximum mean germination time was noticed in  $T_0$  (4.40 days) followed by  $T_{12}$ . Though significant difference was observed in mean germination time but some non-significant difference was also noticed in between  $T_1$ ,  $T_4$ ,  $T_5$ ,  $T_6$  and  $T_{12}$ ,  $T_1$ ,  $T_4$ ,  $T_5$ ,  $T_6$  and  $T_9$ ,  $T_2$ ,  $T_7$ ,  $T_8$  and  $T_{10}$ ,  $T_3$ ,  $T_7$ ,  $T_8$ ,  $T_{10}$  and  $T_{11}$ ,  $T_0$  and  $T_{12}$  (Table 2). Similar kind of experiment was noted by Ray and Bordolui (2022a).

# Germination energy

Among the priming treatments, with different duration and concentration of PEG-6000, significant difference was observed in germination energy. The maximum energy of germination was recorded in  $T_8$  (77.82) followed by  $T_2$  and  $T_{11}$  while it was minimum for  $T_0$  (51.57) preceded by  $T_{12}$  and  $T_6$ . Seed priming treatments enhanced the energy of germination over that of untreated seeds and maximum energy of germination was recorded with hydro-priming in rice (Mahajan *et al.* 2011). Some non-significant difference was also observed in between  $T_1$ ,  $T_3$  and  $T_4$ ,  $T_5$ ,  $T_9$  and  $T_{11}$ ,  $T_6$  and  $T_{12}$  and  $T_2$  and  $T_8$  (Table 2). Sohail

*et al.* (2018) observed similar type of experiment in Kabuli chickpea.

### CONCLUSION

Seeds of chickpea were treated with various concentration and duration of Polyethylene Glycol recorded higher seed quality parameters compared to control. For soaking duration 8 hrs and concentration 20 g liter<sup>-1</sup> of PEG 6000 showed significant performance for seed quality parameter like germination percentage (93.77), seedling fresh weight (1.66 g), seedling dry weight (0.14 g), seedling vigour index I (2137.07), time to 50% germination (2.33 days) and germination index (43.10). Therefore, pre-sowing treatment with PEG 6000 @20 g liter<sup>-1</sup> with duration of 8 hrs is recommended for treating chickpea seed for better seedling establishment.

### REFERENCES

- Abdul Baki AA, Anderson JD (1973) Vigour determination in soybean seed by multiple criteria 1. Crop Science 13 (6): 630—633.
- Amzallag GN, Lerner HR, Poljakoff-Mayber A (1990) Exogenous ABA as a modulator of the response of sorghum to high salinity. J Exp Bot 54 : 1529—1534.
- Arumuganathan K, Earle ED (1991) Nuclear DNA content of some important plant species. *Pl Molecular Biology Reporter* 9 (3): 208–218.
- Association of Official Seed Analysis (AOSA) (1990) Rules for testing seeds. Journal Seed Technology 12: 1112.
- Cayuela E, Perez-Alfocea A, Caro M, Bolarin MC (1996) Priming of seeds with NaCl induces physiological changes in tomato plants grown under salt stress. *Physiol Plant* 96 : 231–236.
- Chakraborty A, Bordolui SK (2021) Impact of seed priming with Ag-Nanoparticle and GA<sub>3</sub> on germination and vigour in green gram. *Int J Curr Microbiol Appl Sci* 10 (03) : 1499—1506 Doi: https://doi.org/10.20546/ijc mas.2021.1003.119.
- Choudhury A, Bordolui SK (2022a) Seed invigoration treatment with sodium molybdate (Na<sub>2</sub>M<sub>o</sub>O<sub>4</sub>) Nutri-priming for improvement of quality performance of Bengal gram (*Cicer arietinum* L.). *The Pharma Innovation Journal* 11 (12): 3381— 3386.
- Choudhury A, Bordolui SK (2022b) Inducement of seed priming with potassium nitrate on quality performance of chickpea (*Cicer arietinum* L.). Biological Forum – An *International Journal* 14 (4) : 779–783.
- Coolbear P, Francis A, Grierson D (1984) The effect of low temperature pre-sowing treatment under the germination performance and membrane integrity of artificially aged tomato

seeds. Journal Exp Botany 35: 1609-1617.

- Dezfuli PM, Zadeh FS, Janmohammadi M (2008) Influence of priming techniques on seed germination behavior of maize inbred lines (*Zea mays L.*). ARPN J Agricult Biolog Sci 3 (3): 22—25.
- Ellis RA, Roberts EH (1981) The quantification of ageing and survival in orthodox seeds. *Seed Sci Technol* 9 : 373–409.
- Farooq M, Basra SMA, Hafeez K, Ahmad N (2005) Thermal hardening : A new seed vigour enhancement tool in rice. Acta Botanical Sinica 47: 187—192.
- Farooq M, Basra SMA, Rehman H, Saleem BA (2008) Seed priming enhances the performance of late sown wheat (*Triticum aestivum* L.) by improving the chilling tolerance. *Journal of Agronomy Crop Sciences* 194: 55—60.
- Ghassemi-Golezani K, Japparpour-Bonyadi Z, Shafagh-Kolvanagh J, Nikpour-Rashidabad N (2013) Effects of water stress and hydro-priming duration on field performance of lentil. *International J Farming Allied Sciences* 2:922– 925.
- Iqbal MA (2015) Improving germination and seedling vigor of cowpea (*Vigna unguiculata* L.) with different priming techniques. *American-Eurasian J Agric Environ Sci* 15 (2): 265—270.
- ISTA (1996) International rules of seed testing. *Rules Seed Science and Technology* 24 (supple) : 1—86.
- Jame YW, Cutforth HW (2004) Simulating the effects of temperature and seeding depth on germination and emergence of spring wheat. *Agricultural and Forest Meteorology* 124 : 207–218.
- Kaur S, Gupta A, Kaur N (2001) Effect of osmo- and hydropriming of chickpea seeds on seedling growth and carbohydrate metabolism under water deficit stress. *Plant Growth Regulation* 37 : 17—22.
- Kaur S, Gupta AK, Kaur N (2000) Effect of GA<sub>3</sub>, kinetin and indole acetic acid on carbohydrate metabolism in chickpea seedlings germinating under water stress. *Pl Growth Regul*

30:61-70.

- Mahajan G, Sarlach RS, Japinder S, Gil MS (2011) Seed priming effects on germination, growth and yield of dry direct-seeded Rice. *Journal of Crop Improvement* 25 (4): 409–417.
- Mohammadi GR (2009) The effect of seed priming on plant traits of late-spring seeded soybean (*Glycine max* L.). Am Eur J Agric Environ Sci 5: 322–326.
- Ozbingol N, Corbineau F, Come D (1998) Response of tomato seeds to osmoconditioning as related to temperature and oxygen. *Seed Sci Res* 8 : 377–384.
- Poehlman JM, Sleper DA (1995) Breeding field crops. 3rded. Iowa state of university press, Iowa 50014.USA.
- Ray J, Bordolui SK (2022a) Effect of seed priming as pre-treatment factors on germination and seedling vigour of tomato. *International Journal Plant Soil Science* 34 (20): 302–311.
- Ray J, Bordolui SK (2022b) Effect of seed priming as pre-treatment factors on germination and seedling vigour of tomato. *International Journal of Plant Soil Science* 34 (20) : 302—311.
- Ruan S, Xue Q, Tylkowska K (2002) Effects of seed priming on germination and health of rice (*Oryza sativa* L.) seeds. *Seed Science Technology* 30 : 451—458.
- Saglam S, Dayi S, Kaya G, Gurbuz A (2010) Hydropriming increases germination of Lentil (*Lens culinaris* Medik.) under water stress. *Not Sci Biol* 2 : 103—106.
- Singh BG, Rao G (1993) Effect of chemical soaking of sunflower (*Helianthus annuus* L.) seed on vigour index. *Indian Journal* of Agricultural Science 63 : 232–233.
- Sohail S, Reddy B, Raj G, Praveena R (2018) Effect of different priming methods on root nodulation in Kabuli chickpea (*Cicer kabulim* L.) seeds. Journal Pharmacognosy and Phytochemistry 7 (4): 2890–2893.
- Srinivasan A, Johansen C, Saxena NP (1998) Cold tolerance during early reproductive growth of chickpea (*Cicer arietinum* L.). Characterization of stress and genotypic variation in pod set. *Field Crops Research* 57 : 181–193.